

RESEARCH

Open Access



# Polymorphisms of the sodium voltage-gated channel, alpha subunit 1 (*SCN1A* -A3184G) gene among children with non-lesional epilepsy: a case-control study

Esraa Ghazala<sup>1</sup>, Doaa A. Shahin<sup>2</sup> and Yahya Wahba<sup>1\*</sup>

## Abstract

**Background:** Mutations in the neuronal sodium voltage-gated channel, alpha subunit 1 (*SCN1A*) gene have been associated with epilepsy. We investigated the *SCN1A*-A3184G polymorphism among Egyptian children and adolescents with non-lesional epilepsy.

**Methods:** A prospective case – control observational study was done in Mansoura University Children's Hospital, Egypt including 326 children with non-lesional epilepsy (163 antiepileptic drugs (AEDs) resistant cases & 163 AEDs responders) and 163 healthy controls. One step real time polymerase chain reaction (PCR) was used for the molecular analysis. Student's t-test, and Monto Carlo, chi-square and Mann–Whitney tests were used for the statistical analysis.

**Results:** All study participants were matched as regards the age, sex and body weight ( $p = 0.07$ ,  $0.347$  and  $0.462$ , respectively). They had the (AA) and (AG) genotypes but not the (GG) variant. No significant differences were found between cases and controls regarding (AG) and (AA) genotypes and A- and G-alleles ( $p = 0.09$  and  $0.3$ , respectively). We did not find significant differences between AEDs responders and resistant cases regarding the studied genotypes and alleles ( $p = 0.61$  and  $0.746$ , respectively). In the resistant group, we observed significant associations between the (AG) genotype and seizure frequency ( $p = 0.05$ ), the tonic-clonic seizure ( $p < 0.001$ ), the younger age of first seizure attack ( $p = 0.03$ ), abnormal electroencephalogram (EEG) ( $p < 0.001$ ), the positive family history of epilepsy ( $p = 0.006$ ), topiramate ( $p = 0.03$ ) and valproic acid ( $p < 0.001$ ), while the (AA) genotype was associated with carbamazepine ( $p = 0.03$ ). While in AEDs responders, there were significant associations between the AG genotype and the abnormal EEG activity, levetiracetam and carbamazepine ( $p = 0.016$ ,  $0.028$  and  $0.02$ ).

**Conclusions:** The *SCN1A*-A3184G genotypes and alleles were not associated with the epilepsy risk among Egyptian children. Significant associations were reported between the AG genotype and some predictors of refractory epilepsy.

**Keywords:** Children, Epilepsy, PCR, *SCN1A* gene

## Background

Epilepsy is a multifactorial channelopathy disease with the involvement of both acquired and genetic factors [1]. Knowing the exact etiology of epilepsy is crucial for

patients' treatment and for neurobiological researches that could direct future personalized therapies [2]. Neuronal voltage-gated sodium channels (SCN) are proteins responsible for the generation and propagation of the action potentials within the neurons. This occurs through affection of the membrane permeability to sodium ions and facilitation of the ions diffusion down

\*Correspondence: wahbayahya2007@mans.edu.eg; yahyawahba@gmail.com

<sup>1</sup> Department of Pediatrics, Mansoura University Faculty of Medicine, Mansoura, Egypt  
Full list of author information is available at the end of the article



an electrochemical gradient till the sodium equilibrium potential [3].

There are evidences about the role of the *SCN* polymorphisms in the epilepsy pathogenesis. These polymorphisms have been associated with a spectrum of epilepsy syndromes such as generalized epilepsy with febrile seizures plus, borderline severe myoclonic epilepsy of infancy, Dravet syndrome, Doose syndrome and infantile spasms [4, 5]. Most of *SCN* mutations are within *SCN1A* gene and fewer are within the *SCN2A*, *SCN8A* and *SCN1B* genes [6]. The *SCN1A* gene is located in the chromosome 2 (2q24.3) and includes 26 exons. *SCN1A* encodes the alpha subunit of the sodium channel NaV1.1 [7]. This type of channels conduct sodium ions through pores in the cellular membranes. Its architecture comprises four domains with six segments; four homologous domains (D1–D4), each containing six  $\alpha$ -helical segments (S1–S6). The positively charged residues in the S4 segment or voltage-sensing helix of each domain are involved in the the graded membrane potential changes and generation of the action potentials [8].

Several studies have reported the efficacy of the sodium-channel blockers for the treatment of epilepsies due to genetic channelopathies. Carbamazepine, oxcarbazepine, phenytoin, lamotrigine, lacosamide and lidocaine prevent seizure activity through blocking the movement of sodium ions across sodium ion channels during the propagation of action potentials. This group of antiepileptic drugs (AEDs) has an additional advantage through acting on potassium channels, due to structural similarities, thus controlling genetic epilepsies due to mutations of voltage-gated potassium channel genes [9].

The *SCN1A-A3184G* (*p.Thr1067Ala*) polymorphism has been suggested to be involved in the gating of sodium channels, thus rendering them insensitive to sodium-channel blockers [10, 11]. The association of the *SCN1A-A3184G* polymorphism with the epilepsy risk was investigated in several non-Caucasian populations [12–15] and in a limited number of Caucasian populations [10, 16] with inconsistent results, thus necessitating additional studies.

Some studies reported significant associations between the *SCN1A* polymorphisms and AEDs resistance [8]. Others tried to investigate the possible associations between these polymorphisms and AEDs choice and doses. Tate et al. reported a significant association between the *SCN1A* polymorphism and the dose of phenytoin and carbamazepine, and suggested a trend of reduction in the maximum dose required according to the genotype [17]. Heinzen et al. noted that individuals with the AA genotype of the *rs3812718* variant need higher doses of AEDs than others with the GG genotype [18].

In the current research, we investigated the patterns and frequencies of *SCN1A-A3184G* (*p.Thr1067Ala*) polymorphism among Egyptian children and adolescents with non-lesional epilepsy, including both AEDs responders and resistant cases. We tested the association between *SCN1A-A3184G* genotypes and some predictors of refractory epilepsy. We also highlighted the possible pharmacological implication of studying the target polymorphism, through describing the frequently used AEDs and their possible link with the genotypes.

## Methods

### Study design and participants

We conducted a preliminary prospective case – control study in Mansoura University Childrens’s Hospital, Mansoura, Egypt from February 2020 to January 2022. We enrolled 326 children with non-lesional epilepsy; 163 AEDs resistant patients and 163 AEDs responders. We included 163 healthy children of comparable genders and ages as a control group. We recruited the controls from the same hospital while they attended for regular follow-up visits or for minor complaints such as pharyngitis and mild gastroenteritis. All controls had no history of neurological disorders, and were of the same ethnic origin as the patients.

Non-lesional epilepsy was described if there was a history of at least two unprovoked seizures accompanied by epileptiform electroencephalogram (EEG) changes in a patient with normal neurological examination and development, and no structural lesions detectable by magnetic resonance imaging [19]. AEDs resistance was defined if at least four seizure attacks happened over 1 year after using the maximum tolerated doses of three appropriate AEDs [20, 21]. Recovery from fits for at least 1 year after starting AEDs was the hallmark for considering the child as an AEDs responder [20, 22]. Patients with a single episode of seizure, metabolic derangement, secondary epilepsies and poor compliance to therapy were excluded.

### Clinical data collection

We retrieved relevant data of the patients from archive files including their gender, age, the seizure type and frequency, the age of onset of seizures, the duration since the last seizure attack (in months), EEG findings and AEDs therapies. We assessed all patients for the seizure type (tonic, tonic-clonic, clonic, myoclonic and absence) using the 2017 International League Against Epilepsy (ILAE) operational classification [19].

### Molecular analysis of the *SCN1A* (*rs2298771*) polymorphism

We extracted genomic DNA from the whole venous blood using QIAamp DNA blood mini kits (provided by

QIAGEN, USA, Catalog Number: 51104) and stored at  $-20^{\circ}\text{C}$  till used. We measured the genomic DNA purity and concentration, from the controls and cases, using the NanoDrop™ 2000 Spectrophotometer (Thermo Scientific, Waltham, MA).

The genotyping of the *SCN1A* (*rs2298771*) polymorphism was carried out by the TaqMan single nucleotide polymorphism (SNP) Genotyping Assays (Applied Biosystem, Foster City, CA). The PCR primers and the variant type allele-specific TaqMan MGB probes were designed by Applied Biosystem. The SNP ID was C 11748767\_20 for the *SCN1A* (*rs2298771*) polymorphism; and the chromosomal location was Chr.2:166036278.

The context sequence [VIC / FAM] was: TAGTCAAGA TCTTTCCCAATTTCTG[C/T]TGTATGATTGGACAT ACAACTGTCT.

As a reporter at the 50 end of TaqMan MGB probe, VIC and FAM were used for the A-allele (Allele-1) and the G-allele (Allele-2), respectively.

We prepared 25  $\mu\text{L}$  of the PCR reaction mix for qPCR analysis as follow: 7.25  $\mu\text{L}$  DNase-free and RNase-free water, 12.5  $\mu\text{L}$  TaqMan universal master mix II with UNG 2 $\times$  (# 4440042, Applied Biosystem), 4  $\mu\text{L}$  DNA template and 1.25  $\mu\text{L}$  TaqMan assay 20 $\times$ . We transferred PCR reaction mix to the 48-well reaction plate that was sealed using an appropriate cover. Then, centrifugation was done, and followed by loading into the step one real-time PCR (Applied Biosystem). The cycling stages of the reaction were as follow: stage I for UNG incubation for 30s at  $60^{\circ}\text{C}$ , stage II for polymerase activation for 10 min at  $95^{\circ}\text{C}$ , stage III for PCR and included 40 cycles of denaturation at  $95^{\circ}\text{C}$  for 15s, annealing/extension at  $60^{\circ}\text{C}$  for 60s and finally stage IV for 30s at  $60^{\circ}\text{C}$ . We

discriminated the alleles by assessing the fluorescence intensity at the endpoint.

We analyzed the results of the measurements using the SDS software version 1.7 (Applied Biosystem), and detected the genotypes (Figs. 1 and 2). We randomly assayed 10% of the original specimens for replicate to exclude errors of genotyping. We did not detect discrepancies between the genotypes which were determined in duplicate.

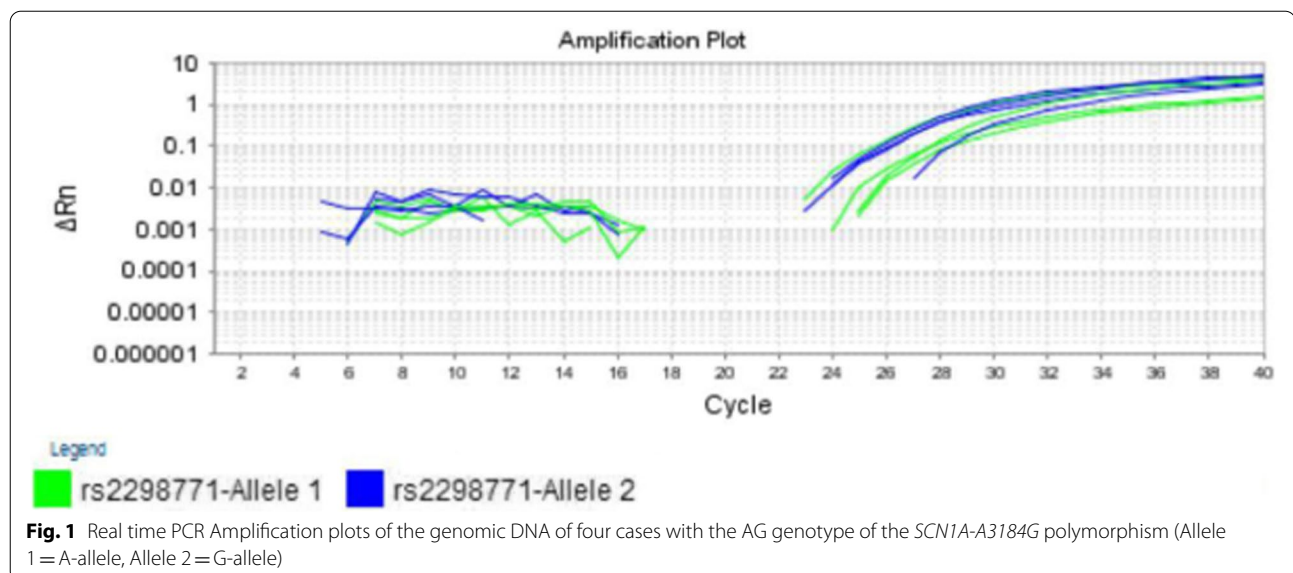
### Statistical analysis

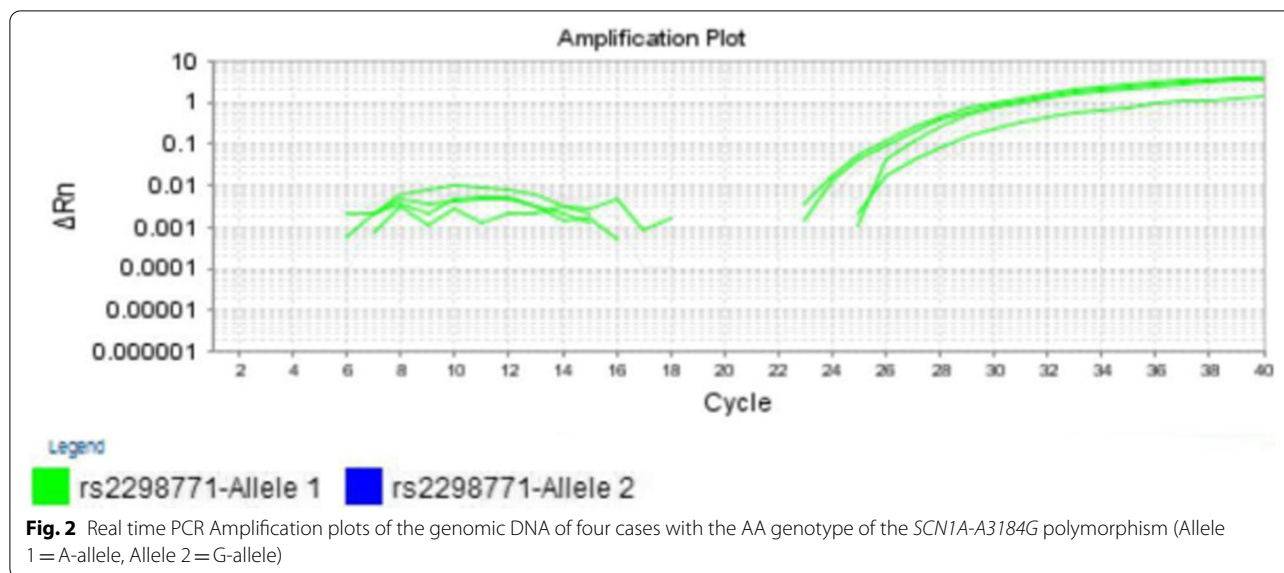
We analyzed data using the Statistical Package for the Social Sciences (SPSS) Version 25 (IBM Corp, Armonk, NY). Numbers (percent) were compared by Monto Carlo and chi-square tests ( $\chi^2$ ). Parametric data (mean  $\pm$  SD) were analyzed by the Student's t-test while non-parametric data (median and interquartile range) were analyzed by the Mann–Whitney test.  $P$  values  $\leq 0.05$  were considered statistically significant. Genotypes and allelic frequencies of *SCN1A* (*rs2298771*) polymorphism were tested using the chi-square test to verify the agreement with Hardy–Weinberg equilibrium ( $p > 0.05$  for each group).

## Results

### Descriptive data and characteristics of the study participants

Both AEDs resistant and AEDs responders groups were matched with the control group as regards the age, gender and body weight ( $p = 0.07$ ,  $0.347$  and  $0.462$ , respectively, Table 1). AEDs-resistant group showed higher seizure frequency, shorter time intervals since the last seizure attack and more abnormal EEG findings ( $p < 0.001$ ,





**Table 1** Characteristics of the study participants

	Control group (n = 163)	AEDs-resistant group (n = 163)	AEDs-responders (n = 163)	p value
Age (years) <sup>a</sup>	9.39 ± 2.27	9.44 ± 3.52	10.17 ± 3.98	0.07
Sex <sup>b</sup>				
Male	78(47.9)	70(42.9)	83(50.9)	0.347
Female	85(52.1)	93(57.1)	80(49.1)	
BW (Kg) <sup>a</sup>	32.78 ± 10	31.46 ± 12.27	36.07 ± 17.93	0.462

Data were presented as mean ± SD<sup>a</sup> and numbers (percent)<sup>b</sup> and analyzed by Student's t-test<sup>a</sup> and chi-square test<sup>b</sup>, respectively  
 AEDs Antiepileptic Drugs, BW Body Weight, Kg Kilogram, N Number, P Probability

Table 2). No statistically significant differences existed between AEDs responders and resistant groups regarding the age of the first seizure attack ( $p = 0.642$ , Table 2). Regarding the type of seizures, the most frequent type of seizures in both groups was the tonic-clonic type that was detected in 63.8 and 76.1% in the AEDs responders and AEDs resistant groups, respectively. The other common types in the AEDs responder group were the tonic seizures (20.9%) and absence seizures (9.2%) while in the AED resistant group, absence seizures were detected in 10.4% and myoclonic seizures in 7.4% (Table 2).

**The pattern of antiepileptic drugs among the patients**

In the current study, valproic acid was the most frequently used AEDs in all groups (82.2% of the AEDs responder group and 90.2% of the AEDs resistant group) followed by levetiracetam in the AEDs resistant group (41.7%) and carbamazepine in the AEDs responder group (27%). There was a statistically significant difference between the

both groups regarding the use of AEDs. Carbamazepine was more frequently used in the AEDs responder group ( $p = 0.03$ ), while levetiracetam was more frequently used in the AEDs resistant group ( $p = 0.003$ ). Topiramate was only used in the AEDs resistant group (Table 2).

**The distribution of SCN1A-A3184G genotypes and alleles among the study groups**

The AG genotype of *SCN1A*-A3184G polymorphism was the predominant genotype in all groups (81.6, 76.1 and 73.6% for the control group, the AEDs responder group, and the AEDs resistant group, respectively, Table 3). No (GG) genotype was described in the studied populations. No significant differences existed between cases and controls regarding (AG) and (AA) genotypes and A- and G-alleles ( $p = 0.09$  and  $0.3$ , respectively).

We did not find significant differences between AEDs responders, AEDs resistant groups and the control group regarding the genotypes and alleles of the *SCN1A*-A3184G polymorphism ( $p = 0.214$  for the genotypes and  $0.563$  for the alleles, Table 3). Moreover, we did not find significant differences between AEDs responders and resistant cases regarding the target genotypes and alleles ( $p = 0.61$  and  $0.746$ , respectively, Table 3).

**Associations between SCN1A-A3184G genotypes and characteristics of the patients**

Regarding combined AEDs responders and resistant cases, Table 4 shows a higher seizure frequency and more generalized epileptiform EEG activity in the AG-genotyped cases ( $p = 0.004$  and  $< 0.001$ , respectively). The frequent seizure types associating the AG genotype were tonic-clonic, tonic and absence seizures, while in patients

**Table 2** Seizure characters and antiepileptic therapies among cases with epilepsy

	AEDs resistant group (n = 163)	AEDs responders (n = 163)	p value
Types of seizures <sup>a</sup>			
Tonic-clonic	124(76.1)	104(63.8)	<0.001*
Tonic	10(6.1)	34(20.9)	
Myoclonic	12(7.4)	0	
Clonic	0	10(6.1)	
Absence	17(10.4)	15(9.2)	
Electroencephalogram <sup>a</sup>			
Normal	33(20.2)	73(44.8)	<0.001*
Generalized activity	130(79.8)	90(55.2)	
Antiepileptic drugs <sup>a</sup>			
Topiramate	12(7.4)	0	<0.001*
Carbamazepine/ Oxcarbazepine	28(17.2)	44(27)	0.03*
Valproic acid	147(90.2)	134(82.2)	0.037*
Levetiracetam	68(41.7)	43(26.4)	0.003*
Polytherapy	86(59.7)	58(40.3)	0.002*
Monotherapy	77(42.3)	105(57.7)	
Duration since the last seizure attack (months) <sup>b</sup>	3(1–3)	14(12–24)	<0.001*
Age of first seizure attack (years) <sup>b</sup>	3(0.75–6)	3(0.83–6)	0.642
Frequency of seizures per month <sup>b</sup>	2(1–4)	0.17(0.17–0.17)	0.001*

Data were expressed as numbers <sup>a</sup> (percent) and median <sup>b</sup> (interquartile range), and tested using chi-square <sup>a</sup> and Monto Carlo tests <sup>a</sup>, and Mann-Whitney test <sup>b</sup>

AEDs Antiepileptic Drugs, N Number, P Probability

\*  $P \leq 0.05$  is significant

**Table 3** Distribution of the *SCN1A-A3184G* genotypes and alleles among the study groups

	Control group (n = 163)	AEDs-resistant (n = 163)	AEDs- responders (n = 163)	p values	OR (95%CI)
Genotypes (n)					
AG	133(81.6)	120(73.6)	124(76.1)	0.214	$p_1 = 0.08$ 1.59(0.94–2.69)
AA	30(18.4)	43(26.4)	39(23.9)		$p_2 = 0.223$ $p_3 = 0.61$ 1.39(0.816–2.38) 0.88(0.53–1.44)
Alleles (2n)					
A	193(59.2)	206(63.2)	202(61.9)	0.563	$p_1 = 0.296$ 0.845(0.617–1.16)
G (r)	133(40.8)	120(36.8)	124(38.04)		$p_2 = 0.471$ $p_3 = 0.746$ 0.891(0.651–1.22) 1.05(0.767–1.45)

Data were presented as numbers (percent) and analyzed by the chi-square test

AEDs Antiepileptic Drugs, CI Confidence Interval, N Number, OR Odds Ratio, P Probability,  $P_1$  for controls versus AEDs-resistant group,  $P_2$  for controls versus AEDs-responders,  $P_3$  for AEDs-resistant group versus AEDs- responders

with the AA genotype were tonic-clonic and tonic seizures. No significant differences existed between the AG and AA genotypes regarding AEDs, except for topiramate that was only used in cases with the AG genotype.

In the AEDs resistant cases, there were significant associations between the AG genotype and the earlier age of onset, the seizures frequency, the abnormal EEG activity, the positive family history of epilepsy, topiramate and valproic acid ( $p = 0.03, 0.05, <0.001, 0.006, 0.03$  and  $<0.001$

respectively, Table 4). While in AEDs responders, there were significant associations between the AG genotype and the abnormal EEG activity, levetiracetam and carbamazepine ( $p = 0.016, 0.028$  and  $0.02$ , respectively, Table 4).

## Discussion

Epilepsy is a highly heterogeneous disease with a confirmed genetic background [23, 24]. More than one quarter of the epilepsy-related genes encode ion channel

**Table 4** Associations between the *SCN1A-A3184G* genotypes and characteristics of the patients

Characteristics	Epilepsy Cases		AEDs resistant group		AEDs responders		p value
	AG(n = 244)	AA(n = 82)	AG(n = 120)	AA(n = 43)	AG(n = 124)	AA (n = 39)	
Family history <sup>a</sup>							$p_1 = 0.268$
Negative	150(61.5)	56(68.3)	60(50)	32(74.4)	90(72.6)	24(61.5)	$p_2 = 0.006^*$
Positive	94(38.5)	26(31.7)	60(50)	11(25.6)	34(27.4)	15(38.5)	$p_3 = 0.19$
Types of seizures <sup>a</sup>							
Tonic-clonic	176(72.1)	52(63.4)	97(80.8)	27(62.8)	79(63.7)	25(64.1)	$p_1 = 0.002^*$
Tonic	25(10.2)	19(23.2)	0	10(23.3)	25(20.2)	9(23.1)	$p_2 < 0.001^*$
Myoclonic	6(2.5)	6(7.3)	6(5)	6(14)	0	0	$p_3 = 0.262$
Clonic	10(4.1)	0	0	0	10(8.1)	0	
Absence	27(11.1)	5(6.1)	17(14.2)	0	10(8.1)	5(12.8)	
EEG <sup>a</sup>							
Normal	60(24.6)	46(56.1)	11(9.1)	22(51.2)	49(39.5)	24(61.5)	$p_1 < 0.001^*$
Generalized epileptiform activity	184(75.4)	36(43.9)	109(90.9)	21(48.8)	75(60.5)	15(38.5)	$p_2 < 0.001^*$ $p_3 = 0.016^*$
AEDs <sup>a</sup>							
Topiramate	12(4.9)	0	12(10)	0	0	0	$p_1 = 0.04^*$ $p_2 = 0.03^*$
Carbamazepine /Oxcarbazepine	50(20.5)	22(26.8)	11(9.2)	17(39.5)	39(31.5)	5(12.8)	$p_1 = 0.231$ $p_2 = 0.03^*$ $p_3 = 0.02^*$
Levetiracetam	86(35.2)	25(30.5)	48(40)	20(46.5)	38(30.6)	5(12.8)	$p_1 = 0.432$ $p_2 = 0.457$ $p_3 = 0.028^*$
Valproic acid	215(88.1)	66(80.5)	115(95.8)	32(74.4)	100(80.6)	34(87.2)	$p_1 = 0.083$ $p_2 < 0.001^*$ $p_3 = 0.352$
Duration since the last seizure (months) <sup>b</sup>	3(0.75–5)	3(0.94–9)	2(1–6)	3(0.75–3)	18(12–24)	13(12–24)	$p_1 = 0.088$ $p_2 = 0.029^*$ $p_3 = 0.139$
Age of the first seizure attack (years) <sup>b</sup>	2(1–5)	1(0.33–3)	3(0.75–6)	3(1–9)	3(1–5)	2.5(0.67–8)	$p_1 = 0.015^*$ $p_2 = 0.03^*$ $p_3 = 0.229$
Frequency of seizures (per month) <sup>b</sup>	13(12–24)	12(6–24)	2(1–5)	1(0.33–4)	0	0.17(0.17–0.17)	$p_1 = 0.004^*$ $p_2 = 0.05^*$

Data were expressed as numbers <sup>a</sup> (percent) and median <sup>b</sup> (interquartile range), and compared using the chi-square <sup>a</sup> and Monto Carlo tests <sup>a</sup>, and Mann-Whitney test <sup>b</sup>

AEDs Antiepileptic Drugs, Epilepsy cases AEDs responders plus resistant cases, EEG Electroencephalogram, N Number, P Probability,  $P_1$  for the AG versus AA genotypes in epilepsy cases,  $P_2$  for the AG versus AA genotypes in the AEDs-resistant group,  $P_3$  for the AG versus AA genotypes in AEDs-responders

\*  $P \leq 0.05$  is significant

proteins, including the ligand-gated ion channels (such as gamma-aminobutyric acid receptors, N-methyl-D-aspartate receptors and nicotinic acetylcholine receptors) and the voltage-gated channels (such as  $Ca^{2+}$ ,  $K^+$  and  $Na^+$  channels) [25]. The first *SCN1A* mutation was found in epilepsy patients by 2000 [23], but now many new *SCN1A* mutations have been identified making it the most common epilepsy-related gene [26]. In our preliminary research, we investigated the patterns and frequencies of *SCN1A-A3184G* alleles and genotypes among Egyptian children and adolescents with non-lesional epilepsy. We found that the AG genotype and the A allele are the predominant models among the study participants.

In the current study, we reported insignificant differences between epilepsy cases and the control group regarding the *SCN1A-A3184G* genotypes and alleles. No significant differences existed between AEDs responders and resistant cases regarding these genotypes and alleles. Lack of significant differences among our Egyptian patients could be explained on the basis of the ethnic variation. This was evidenced by Baum et al. in their large multi-ethnic study, where they considered the *SCN1A-A3184G* (*rs2298771*) polymorphism as the significant ethnic-related locus among symptomatic epilepsy patients ( $p < 0.001$ ) while the frequencies of the other studied *SCN1A* loci (*rs10188577* and *rs3812718*) did not

differ according to the ethnicity [15]. However, the relatively small sample size could be a limiting factor in interpreting our results.

Our results also agree with Chou et al. who reported insignificant differences between 83 control subjects and 104 Taiwanese epileptic children regarding *SCN1A-A3184G* genotypes and alleles [14]. We also agree with Kang et al. who investigated 311 children for *SCN1A-A3184G* (*rs2298771*) and *SCN2A-G56A* (*rs17183814*) polymorphisms and tested their association with refractory seizures. They reported insignificant differences between controls and cases as regards the *SCN1A-A3184G* polymorphism, but the A allele of *SCN2A-G56A* polymorphism was associated with the refractory seizures [27]. Moreover, similar findings were described in a German study where *SCN1A-A3184G* polymorphism was not considered as a major contributor to the idiopathic generalized epilepsy [28]. We also agree with Hosseini et al. who evaluated the frequency of the *SCN1A* (*rs2298771* & *7,601,520*) and the *ABCB1* (*rs1045642*) polymorphisms within the Iranian population with idiopathic refractory epilepsy. They enrolled 81 healthy subjects and 34 patients with idiopathic refractory epilepsy, and reported insignificant differences between the studied groups [29]. There is also agreement with another Chinese study including 471 patients with epilepsy (272 AEDs responders and 199 AEDs resistant cases). Authors studied the association of the responsiveness to AEDs with *SCN1A*, *SCN2A*, and *SCN3A* polymorphisms, and correlated any association with the mRNA expression of the neuronal sodium channels. They suggested an association between *SCN2A IVS7-32A >G* and the AEDs response, without any evidence of an effect on the mRNA expression or splicing. However, they reported a negative association between the *SCN1A-A3184G* polymorphism and the resistance to AEDs [30].

On the other hand, Lakhan et al. suggested that the AG genotype of the *SCN1A-A3184G* polymorphism was more frequent among North Indian epilepsy patients [ $p=0.005$ ; 95% confidence interval=1.19–2.61, odds ratio=1.76] [12]. In a meta-analysis study done by Li et al., the *SCN1A-A3184G* (*rs2298771*) polymorphism was significantly associated with the response to AEDs [31].

In the current study, we did not find the GG genotype of the *SCN1A-A3184G* polymorphism among the studied population. This is in contrast to a Slovenian study where the G allele was associated with a lower epilepsy risk and a high remission rate among patients with epilepsy [7]. These discrepancies could be mainly explained by the interethnic differences in the *SCN1A* alleles/genotypes distribution [20].

Interestingly, the current study suggested significant associations between the heterozygous AG genotype in the epilepsy cases with the higher seizure frequency and

generalized epileptiform EEG activity. Possible associations were also suggested between the AG-genotyped patients with refractory epilepsy and the earlier age of onset of seizures and duration since the last seizure attack. On reviewing literature, no previous reports tested the associations between the *SCN1A-A3184G* (*rs2298771*) polymorphism and the predictors of refractory epilepsy. We would like to emphasize that the higher seizure frequency, generalized epileptiform EEG activity and the earlier age of onset of seizures were proved to be significant predictors of the AEDs response in previous studies [32, 33]. This observation could open the gate for considering the AG genotype of the *SCN1A-A3184G* polymorphism as a predictor for treatment response, and aid in designing the personalized medicine for epileptic children according to their genotype [34]. However, large scale multi-center studies are still needed to prove such associations.

Several studies were carried out to select the most appropriate AEDs in different types of epilepsy [9, 35]. The current study suggested that levetiracetam and carbamazepine were more frequently used in the AG-genotyped responders than those with the AA genotype. Regarding the AEDs resistant group, topiramate and valproic acid were more frequently used in children with the AG genotype while carbamazepine was more frequently used in the AA genotype. This observation highlights the possible role of the *SCN1A-A3184G* genotypes in the selection of the appropriate AEDs.

On reviewing literature, several studies were carried out to detect the possible pharmacological implications of the *SCN1A-A3184G* polymorphism in epilepsy management. A recent meta-analysis including eight studies reported that Asian patients with epilepsy and the GG genotype of the *SCN1A-A3184G* polymorphism are at a higher risk of carbamazepine resistance [36]. On the other hand, another meta-analysis including 18 studies (2546 patients) reported no association between the same polymorphism and the carbamazepine resistance [37]. Regarding the valproic acid, a Chinese study suggested that *SCN1A-A3184G* polymorphism has no effect on the drug response [38]. Actually, gathering data from different ethnic groups expand the knowledge about the genetic background of epilepsy and help in personalized medicine strategy.

#### Study limitations

A single center study, the lack of serum AEDs levels assay and limited studied variables.

#### Conclusions

No significant differences were found between Egyptian children and adolescents with non-lesional epilepsy and healthy controls regarding the frequency of

**SCN1A-A3184G polymorphism.** We suggested significant associations between the AG genotype of the *SCN1A-A3184G* polymorphism and some predictors of refractory epilepsy. The *SCN1A-A3184G* genotypes might affect AEDs selection. Multicenter large-scale studies are still needed to validate our findings.

#### Abbreviations

AEDs: Antiepileptic Drugs; EEG: Electroencephalogram; PCR: Polymerase Chain Reaction; SCN: Sodium Channels; SCN1A: Sodium Channel, Alpha Subunit 1; SNP: Single Nucleotide Polymorphism; SPSS: Statistical Package for the Social Sciences.

#### Acknowledgements

We greatly appreciate the role of professor Mostafa El Ayouty, the former head of the pediatric neurology unit of Mansoura University Faculty of Medicine, for his help and support in the access to patients and data.

#### Authors' contributions

All authors contributed to the study design and conception. Preparation of materials, data collection and analysis were carried out by EG and YW. The molecular work-up was done by DAS. The first draft of the manuscript was written by YW, and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

#### Funding

Open access funding provided by The Science, Technology & Innovation Funding Authority (STDF) in cooperation with The Egyptian Knowledge Bank (EKB). No funds, grants, or other support was received.

#### Availability of data and materials

The datasets generated during the current study are available in the Mendeley Data: <https://doi.org/10.17632/mkjprdydst.1>

#### Declarations

##### Ethics approval and consent to participate

This study was performed in accordance with the principles of the Declaration of Helsinki, and approved by the Institutional Research Board of Mansoura University Faculty of Medicine, Mansoura, Egypt (Code No: MS.20.02.1041). Written informed consent was obtained from parents and/or legal guardians of the study participants.

##### Consent for publication

Not applicable.

##### Competing interests

The authors declare that they have no conflict of interest.

##### Author details

<sup>1</sup>Department of Pediatrics, Mansoura University Faculty of Medicine, Mansoura, Egypt. <sup>2</sup>Department of Clinical Pathology (Hematology), Mansoura University Faculty of Medicine, Mansoura, Egypt.

Received: 20 July 2022 Accepted: 19 August 2022

Published online: 02 September 2022

#### References

- Helbig I. Genetic causes of generalized epilepsies. *Semin Neurol.* 2015;35(3):288–92.
- Thomas RH, Berkovic SF. The hidden genetics of epilepsy—a clinically important new paradigm. *Nat Rev Neurol.* 2014;10(5):283–92.
- Hille B. Ion channels of excitable membranes. 3rd ed. Sinauer: Sunderland; 2001.
- Harkin LA, McMahon JM, Iona X, Dibbens L, Pelekanos JT, Zuberi SM, et al. The spectrum of SCN1A-related infantile epileptic encephalopathies. *Brain.* 2007;130(Pt 3):843–52.
- Usluer S, Salar S, Arslan M, Yiş U, Kara B, Tektürk P, et al. SCN1A gene sequencing in 46 Turkish epilepsy patients disclosed 12 novel mutations. *Seizure.* 2016;39:34–43.
- Kaplan DI, Isom LL, Petrou S. Role of sodium channels in epilepsy. *Cold Spring Harb Perspect Med.* 2016;6(6):a022814.
- Bertok S, Dolžan V, Goričar K, Podkrajšek KT, Battelino T, Renner-Primec Z. The association of SCN1A p.Thr1067Ala polymorphism with epilepsy risk and the response to antiepileptic drugs in Slovenian children and adolescents with epilepsy. *Seizure.* 2017;51:9–13.
- Catterall WA. Ion channel voltage sensors: structure, function, and pathophysiology. *Neuron.* 2010;67(6):915–28.
- Falsaperla R, Scalia B, Giugno A, Pavone P, Motta M, Caccamo M, et al. Treating the symptom or treating the disease in neonatal seizures: a systematic review of the literature. *Ital J Pediatr.* 2021;47(1):85.
- Abo El Fotoh WM, Abd El Naby SA, Habib MS, ALrefai AA, Kasemy ZA. The potential implication of SCN1A and CYP3A5 genetic variants on antiepileptic drug resistance among Egyptian epileptic children. *Seizure.* 2016;41:75–80.
- Wang P, Zhou Q, Sheng Y, Tang B, Liu Z, Zhou B. Association between two functional SNPs of SCN1A gene and efficacy of carbamazepine monotherapy for focal seizures in Chinese Han epileptic patients. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* 2014;39(5):433–41.
- Lakhan R, Kumari R, Misra UK, Kalita J, Pradhan S, Mittal B. Differential role of sodium channels SCN1A and SCN2A gene polymorphisms with epilepsy and multiple drug resistance in the north Indian population. *Br J Clin Pharmacol.* 2009;68(2):214–20.
- Ebrahimi A, Houshmand M, Tonekaboni SH, Fallah Mahboob Passand MS, Zainali S, Moghadasi M. Two novel mutations in SCN1A gene in Iranian patients with epilepsy. *Arch Med Res.* 2010;41(3):207–14.
- Chou IC, Peng CT, Tsai FJ, Huang CC, Shi YR, Tsai CH. The lack of association between febrile convulsions and polymorphisms in SCN1A. *Epilepsy Res.* 2003;54(1):53–7.
- Baum L, Haerian BS, Ng HK, Wong VC, Ng PW, Lui CH, et al. Case-control association study of polymorphisms in the voltage-gated sodium channel genes SCN1A, SCN2A, SCN3A, SCN1B, and SCN2B and epilepsy. *Hum Genet.* 2014;133(5):651–9.
- Daci A, Beretta G, Vllasaliu D, Shala A, Govori V, Norata GD, et al. Polymorphic variants of SCN1A and EPHX1 influence plasma carbamazepine concentration, metabolism and pharmacoresistance in a population of Kosovar Albanian epileptic patients. *PLoS One.* 2015;10(11):e0142408.
- Tate SK, Depondt C, Sisodiya SM, Cavalleri GL, Schorge S, Soranzo N, et al. Genetic predictors of the maximum doses patients receive during clinical use of the anti-epileptic drugs carbamazepine and phenytoin. *Proc Natl Acad Sci U S A.* 2005;102(15):5507–12.
- Heinzen EL, Yoon W, Tate SK, Sen A, Wood NW, Sisodiya SM, et al. Nova2 interacts with a cis-acting polymorphism to influence the proportions of drug-responsive splice variants of SCN1A. *Am J Hum Genet.* 2007;80(5):876–83.
- Fisher RS, Cross JH, French JA, Higurashi N, Hirsch E, Jansen FE, et al. Operational classification of seizure types by the international league against epilepsy: position paper of the ILAE Commission for Classification and Terminology. *Epilepsia.* 2017;58(4):522–30.
- Eltalal S, El Ayouty M, El-Said A, Wahba Y. CYP2C9 (\*2&\*3) and CYP2C19 (\*2&\*3) polymorphisms among children with nonlesional epilepsy: a single-center study. *Acta Neurol Belg.* 2021;121(6):1623–31.
- Siddiqui A, Kerb R, Weale ME, Brinkmann U, Smith A, Goldstein DB, et al. Association of multidrug resistance in epilepsy with a polymorphism in the drug-transporter gene ABCB1. *N Engl J Med.* 2003;348(15):1442–8.
- Kumari R, Lakhan R, Garg R, Kalita J, Misra U, Mittal B. Pharmacogenomic association study on the role of drug metabolizing, drug transporters and drug target gene polymorphisms in drug-resistant epilepsy in a north Indian population. *Indian J Hum Genet.* 2011;17(Suppl 1):32–40.
- Oyler J, Maljevic S, Scheffer IE, Berkovic SF, Petrou S, Reid CA. Ion channels in genetic epilepsy: from genes and mechanisms to disease-targeted therapies. *Pharmacol Rev.* 2018;70(1):142–73.
- Elsaid AM, Zahran RF, Elmetwaly SM, Wahba Y, Megahed H, Elshazli RM. The potential impact of CYP2D6 (\*2/\*4/\*10) gene variants among Egyptian epileptic children: a preliminary study. *Gene.* 2022;832:146585.
- Catterall WA, Goldin AL, Waxman SG. International Union of Pharmacology. XLVII. Nomenclature and structure-function relationships of voltage-gated sodium channels. *Pharmacol Rev.* 2005;57(4):397–409.



26. Oliva M, Berkovic SF, Petrou S. Sodium channels and the neurobiology of epilepsy. *Epilepsia*. 2012;53(11):1849–59.
27. Kang JW, Kim DW, Lee YS, Lee YH, Lee KS. Polymorphism of SCN1A and SCN2A gene in pediatric refractory epilepsy patients. *J Korean Epilepsy Soc*. 2012;16:49–55.
28. Escayg A, Heils A, MacDonald BT, Haug K, Sander T, Meisler MH. A novel SCN1A mutation associated with generalized epilepsy with febrile seizures plus—and prevalence of variants in patients with epilepsy. *Am J Hum Genet*. 2001;68(4):866–73.
29. Hosseini M, Ebrahimi A, Houshmand M, Zainali S, Tonekaboni SH, Moghaddasi M. SCN1A and ABCB1 polymorphisms in epilepsy. *Arch Neurosci*. 2018;5(1):e59383.
30. Kwan P, Poon WS, Ng HK, Kang DE, Wong V, Ng PW, et al. Multidrug resistance in epilepsy and polymorphisms in the voltage-gated sodium channel genes SCN1A, SCN2A, and SCN3A: correlation among phenotype, genotype, and mRNA expression. *Pharmacogenet Genomics*. 2008;18(11):989–98.
31. Li M, Zhong R, Lu Y, Zhao Q, Li G, Lin W. Association between SCN1A rs2298771, SCN1A rs10188577, SCN2A rs17183814, and SCN2A rs2304016 polymorphisms and responsiveness to antiepileptic drugs: a meta-analysis. *Front Neurol*. 2020;11:591828.
32. Tripathi M, Padhy UP, Vibha D, Bhatia R, Padma Srivastava MV, Singh MB, et al. Predictors of refractory epilepsy in North India: a case–control study. *Seizure*. 2011;20(10):779–83.
33. Fray S, Kchaou M, Chebbi S, Belal S. Predictors factors of refractory epilepsy in childhood. *Rev Neurol (Paris)*. 2015;171(10):730–5.
34. Helbig I, Ellis CA. Personalized medicine in genetic epilepsies—possibilities, challenges, and new frontiers. *Neuropharmacology*. 2020;172:107970.
35. Falsaperla R, Vitaliti G, Mauceri L, Romano C, Pavone P, Motamed-Gorji N, et al. Levetiracetam in neonatal seizures as first-line treatment: a prospective study. *J Pediatr Neurosci*. 2017;12(1):24–8.
36. Zhang X, Liu J, Ye J. Association between SCN1A polymorphism and carbamazepine responsiveness in epilepsy: a meta-analysis. *Epilepsy Res*. 2021;176:106627.
37. Zhao GX, Zhang Z, Cai WK, Shen ML, Wang P, He GH. Associations between CYP3A4, CYP3A5 and SCN1A polymorphisms and carbamazepine metabolism in epilepsy: a meta-analysis. *Epilepsy Res*. 2021;173:106615.
38. Shi L, Zhu M, Li H, Wen Z, Chen X, Luo J, et al. SCN1A and SCN2A polymorphisms are associated with response to valproic acid in Chinese epilepsy patients. *Eur J Clin Pharmacol*. 2019;75(5):655–63.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

