RESEARCH



Chromosome 1p36 candidate gene ZNF436 predicts the prognosis of neuroblastoma: a bioinformatic analysis



Haiwei Wang^{1*†}, Xinrui Wang^{1†} and Liangpu Xu¹

Abstract

Background Genetic 1p deletion is reported in 30% of all neuroblastoma and is associated with the unfavorable prognosis of neuroblastoma. The expressions and prognosis of 1p candidate genes in neuroblastoma are unclear.

Methods Public neuroblastoma cohorts were obtained for secondary analysis. The prognosis of 1p candidate genes in neuroblastoma was determined using Kaplan-Meier and cox regression analysis. The prediction of the nomogram model was determined using timeROC.

Results First, we confirmed the bad prognosis of 1p deletion in neuroblastoma. Moreover, zinc finger protein 436 (ZNF436) located at 1p36 region was down-regulated in 1p deleted neuroblastoma and higher ZNF436 expression was associated with the longer event free survival and overall survival of neuroblastoma. The expression levels of ZNF436 were lower in neuroblastoma patients with MYCN amplification or age at diagnosis ≥ 18months, or with stage 4 neuroblastoma. ZNF436 had robust predictive values of MYCN amplification and overall survival of neuroblastoma. Furthermore, the prognostic significance of ZNF436 in neuroblastoma was independent of MYCN amplification and age of diagnosis. Combinations of ZNF436 with MYCN amplification or age of diagnosis achieved better prognosis. At last, we constructed a nomogram risk model based on age, MYCN amplification and ZNF436. The nomogram model could predict the overall survival of neuroblastoma with high specificity and sensitivity.

Background

Conclusions Chromosome 1p36 candidate gene ZNF436 was a prognostic maker of neuroblastoma.

Keywords Neuroblastoma, 1p deletion, MYCN amplification, ZNF436, Nomogram model

[†]Haiwei Wang and Xinrui Wang equally contributed to this work.

*Correspondence: Haiwei Wang hwwang@sibs.ac.cn

¹Fujian Maternity and Child Health Hospital, Affiliated Hospital of Fujian Medical University, Fuzhou, Fujian, China

The malignancy and clinical outcomes of neuroblastoma are significantly varied [5]. Known prognostic factors of neuroblastoma include age at diagnosis, neuroblastoma stage and MYCN amplification [6]. MYCN maps to the chromosome 2p24.3 region and gain of MYCN copy number variation is detected in about 25% high risk

Neuroblastoma is a common extracranial tumor in children [1, 2]. The clinical outcomes of high risk neuroblastoma are unsatisfied and about 10-15% of pediatric cancer related mortality is associated with neuroblastoma [3, 4]. Neuroblastoma is a heterogeneous disease.

© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

neuroblastoma [3]. The size of MYCN amplicon ranges from 100 to 1500 Kb, including MYCN, DDX1 and some other genes [7, 8]. Except 2p24.3 amplification, overall survival is lower in neuroblastoma patients harboring telomere, RAS and TP53 mutations [9]. Understanding the correlations between genetic aberrations and clinical features of neuroblastoma will provide novel therapeutic targets and prognostic makers for neuroblastoma.

Loss of 1p, 11q and gain of 17q amplification are also well known biomarkers correlated with the clinical risks and outcomes of neuroblastoma [10-12]. Particularly, deletion of 1p36 has been reported in 10-30% of neuroblastoma and is associated with the unfavorable prognosis of neuroblastoma [13, 14]. Chromosome 1p36 regions harbor multiple potential neuroblastoma suppressor genes [15, 16]. For example, CHD5 is located at 1p36.31 region [17, 18] and is associated with the lower overall survival of neuroblastoma [19, 20]. Furthermore, forced expression of CHD5 in neuroblastoma cells with 1p deletion suppresses the metastatic progress of neuroblastoma [21]. TP73 is a TP53 homologue and localized at 1p36.3 region [22]. TP73 could regulate the expressions of MYCN [23] and induce the differentiation of neuroblastoma [24]. Chromosome 1p36.22 candidate gene KIF1B is a tumor suppressor gene in neuroblastoma [25] and germline KIF1B deletion is associated with high predisposition of neuroblastoma development [26]. All those results highlighted the important pathogenic roles of 1p candidate genes in neuroblastoma. However, the expressions and prognosis of neuroblastoma suppressor genes in chromosome 1p regions are not studied in a comprehensive manner.

Using published neuroblastoma cohorts, we analyzed the expressions and prognosis of 1p candidate genes. Our results suggested that zinc finger protein 436 (ZNF436) was an independent prognostic marker and was significantly associated with the favorable clinical outcomes of neuroblastoma. Combinations of ZNF436, MYCN amplification or age at diagnosis achieved better prognosis in neuroblastoma. Finally, we showed that a nomogram risk model based on age, MYCN amplification and ZNF436 could predict the overall survival of neuroblastoma with high specificity and sensitivity.

Methods

Data collection

Therapeutically Applicable Research to Generate Effective Treatments (TARGET) datasets were collected from https://ocg.cancer.gov/ website [27]. The copy number variations of TARGET dataset were downloaded from cBioPortal (http://www.cbioportal.org/) [28, 29]. E-MTAB-1781 dataset [30] was collected from The European Molecular Biology Laboratory-European Bioinformatics Institute (EMBL-EBI) (https://www.ebi.ac.uk/ arrayexpress/). The Gene Expression Omnibus (GEO) datasets GSE13136 [31], GSE73517 [32], GSE16476 [33–35], GSE62564 [36] and GSE85047 [37] were collected from www.ncbi.nlm.nih.gov/geo website. All datasets were analyzed using R software. The expression of genes in neuroblastoma were showed using "pheatmap" package. All the samples used in this study were derived from primary untreated neuroblastoma tumors.

Univariate and multivariate cox regression analysis

Packages "survival" and "survminer" were used for univariate and multivariate cox regression analysis. Packages "forestplot" and "ggforest" were used to generate the forest plots. The Hazard ratio (HR) and P values were determined during cox regression survival analysis.

Kaplan-Meier survival analysis

Kaplan-Meier survival analysis was carried out using "survival" and "survminer" packages. Neuroblastoma patients were classified into "high" or "low" groups by the best cutoff points. P values were determined by log-rank test.

Receiver operating characteristic (ROC) and timeROC analysis

The ROC curves were plotted by 'pROC' package. The timeROC curves were generated using "timeROC" package. The area under the ROC curve (AUC) was determined by the 'pROC' and "survival" packages.

Construction and validation of nomogram model

First, nomogram models were constructed using packages"rms" and "ggplot2" based on age, MYCN amplification and ZNF436 expression levels. Second, the accuracy of the nomogram model was further evaluated using calibration diagram. Third, the risk point of each patient in the nomogram model was determined by "nomogram-Formula". Neuroblastoma patients were classified into higher risk groups or lower risk groups. At last, Kaplan-Meier survival analysis and timeROC analysis were used to validate the prognostic roles of the nomogram models.

Statistical analysis

The statistical P values were performed using two tails paired student's t test. P values less than 0.05 were indicated significant difference.

Results

Genetic 1p deletion is an Independent prognostic marker of neuroblastoma

First, we confirmed the prognosis of 1p deletion in neuroblastoma using TARGET copy number variation dataset. The copy number variation dataset in TARGET only included 59 neuroblastoma patients and only four neuroblastoma patients were with 1p36 deletion. Compared with neuroblastoma patients with normal 1p36, neuroblastoma patients with 1p36 deletion had shorter event free survival (Fig. 1a). Moreover, neuroblastoma patients carrying 1p36 deletion had worse overall survival, contrast with neuroblastoma patients with intact 1p36 in TARGET dataset (Fig. 1a).

The prognosis of 1p deletion was further confirmed using E-MTAB-1781 dataset. There were 151 neuroblastoma patients with 1p deletion, while, 428 neuroblastoma patients were with normal 1p. Similarly, 1p deletion was correlated with the prognosis of neuroblastoma in E-MTAB-1781 dataset. Compared with neuroblastoma patients with normal 1p, neuroblastoma patients with 1p deletion had shorter event free survival and overall survival (Fig. 1b).

Age at diagnosis and MYCN amplification are critical prognostic makers of neuroblastoma [38]. More than 98% neuroblastoma patients with normal 1p were without MYCN amplification, while, 47% neuroblastoma patients with 1p deletion were with amplified MYCN (Fig. 1c). Moreover, neuroblastoma patients with age at diagnosis < 18 months were associated with 1p intact status, while neuroblastoma patients with age at diagnosis > 18 months were associated with 1p deletion status (Fig. 1c). Furthermore, 1p deletion was a prognostic factor of neuroblastoma in E-MTAB-1781, independent of MYCN amplification and age at diagnosis (Fig. 1d).

Expressions and prognosis of 1p candidate genes in neuroblastoma

Genetic 1p regions include multiple tumor suppressor genes. However, not all genes located in 1p were downregulated with the deletion of 1p and associated with the prognosis of neuroblastoma. So, we tried to analyze the expressions and prognosis of 1p candidate genes in neuroblastoma cohorts. Based on TARGET dataset in cBioPortal, 773 genes in chromosomal 1p regions were deleted in neuroblastoma patients. Compared with 1p normal neuroblastoma patients, only 105 genes in 1p regions were differentially expressed in 1p deleted neuroblastoma patients in E-MTAB-1781 dataset (Fig. 2a). Interestingly, those differentially expressed genes in 1p regions were associated with MYCN transcription factor (Fig. 2b).

The differentially expressed genes in 1p deleted neuroblastoma patients in GSE13136 and GSE73517 datasets were also identified. There were 25 and 280 genes in 1p regions were differentially expressed in 1p deleted patients in GSE13136 and GSE73517 datasets, respectively (Fig. 2a). Among them, BCAR3, CLSTN1, CTN-NBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12, UBE2J2 and ZNF436 were commonly changed in 1p deleted neuroblastoma patients in E-MTAB-1781,

Furthermore, we determined the prognostic effects of those commonly down-regulated genes in neuroblastoma patients using univariate cox regression analysis. In E-MTAB-1781 dataset, BCAR3, CLSTN1, CTNNBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12 and ZNF436 were all associated with the favorable prognosis of neuroblastoma. However, UBE2J2 had not prognosis of neuroblastoma (Fig. 2d). The prognostic effects of BCAR3, CLSTN1, CTNNBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12, UBE2J2 and ZNF436 in neuroblastoma patients were further validated using TARGET, GSE16476, GSE62564 and GSE85047 datasets. CTNNBIP1, DNAJC8, NMNAT1, STX12 and ZNF436 represented significantly prognostic factors in both TAR-GET and GSE16476 datasets (Fig. 2d). Only a few differentially expressed genes were detected in GSE62564 and GSE85047 datasets and all those detected genes were associated with the favorable prognosis of neuroblastoma (Fig. 2d). Results from five independent neuroblastoma cohorts suggested that ZNF436 was detected and was associated with the favorable prognosis of neuroblastoma in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets.

High expression levels of ZNF436 are associated with the favorable prognosis of neuroblastoma

ZNF436 is a transcription factor in 1p36.12 regions. The Kaplan-Meier survival analysis was further used to validate the favorable prognosis of ZNF436 in neuroblastoma. Neuroblastoma patients with ZNF436 higher expression levels had prolonged event free survival contrast with ZNF436 lowly expressed neuroblastoma patients in E-MTAB-1781 dataset (Fig. 3a). Also, neuroblastoma patients with ZNF436 higher expression levels had prolonged overall survival contrast with ZNF436 lowly expressed neuroblastoma patients in E-MTAB-1781 dataset (Fig. 3a). Also, neuroblastoma patients with ZNF436 higher expression levels had prolonged overall survival contrast with ZNF436 lowly expressed neuroblastoma patients in E-MTAB-1781 dataset (Fig. 3b).

The prognostic effects of ZNF436 in neuroblastoma were further validated using TARGET, GSE16476, GSE62564 and GSE85047 datasets. Similar with E-MTAB-1781 dataset, neuroblastoma patients with ZNF436 higher expression levels had prolonged event free survival (Fig. 3a) and overall survival (Fig. 3b), contrast with ZNF436 lowly expressed neuroblastoma patients in TARGET, GSE16476, GSE62564 and GSE85047 datasets.

Expressions of ZNF436 in different sub-types of neuroblastoma

Age at diagnosis, MYCN amplification and neuroblastoma stage are critical biomarkers of neuroblastoma



Fig. 1 Prognosis of 1p deletion in neuroblastoma. (a) Event free survival and overall survival of neuroblastoma with 1p deletion or without 1p deletion in TARGET dataset. P values were determined by Log-rank test. (b) Event free survival and overall survival of neuroblastoma with 1p deletion or without 1p deletion in E-MTAB-1781 dataset. (c) Clinical characteristics of neuroblastoma patients in E-MTAB-1781 dataset. (d) Forest plot showed the prognosis of age, MYCN amplification and 1p deletion in E-MTAB-1781 dataset. Hazard ratio (HR) and P values were determined by multivariate cox regression analysis



Fig. 2 Expressions and prognosis of 1p candidate genes in neuroblastoma. (a) The common differentially expressed genes in 1p deleted neuroblastoma in E-MTAB-1781, GSE13136 and GSE73517 datasets. (b) Enriched transcription factors using the differentially expressed genes in 1p deleted neuroblastoma in E-MTAB-1781 dataset. (c) Expressions of BCAR3, CLSTN1, CTNNBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12, UBE2J2 and ZNF436 in neuroblastoma with 1p deletion or without 1p deletion in E-MTAB-1781, GSE13136 and GSE73517 datasets. (d) Forest plots showed the associations of BCAR3, CLSTN1, CTNNBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12, UBE2J2 and ZNF436 with the neuroblastoma overall survival in E-MTAB-1781, GSE13136 and GSE73517 datasets. (d) Forest plots showed the associations of BCAR3, CLSTN1, CTNNBIP1, DNAJC8, HMGCL, NMNAT1, PANK4, PHF13, STX12, UBE2J2 and ZNF436 with the neuroblastoma overall survival in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets. HR and P values were determined by univariate cox regression analysis



Fig. 3 High expression levels of ZNF436 are associated with the favorable prognosis of neuroblastoma. (a) The Kaplan-Meier curves showed the different event free survival of neuroblastoma patients with ZNF436 higher expressions or lower expressions in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets. (b) Different overall survival of neuroblastoma patients with ZNF436 higher expressions or lower expressions or lower expressions in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets. (b) Different overall survival of neuroblastoma patients with ZNF436 higher expressions or lower expressions in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets.

[38]. So, we determined the ZNF436 expressions in different sub-types of neuroblastoma using seven independent neuroblastoma cohorts. ZNF436 was down-regulated in MYCN amplified neuroblastoma patients in E-MTAB-1781, GSE13136, GSE73517, TARGET, GSE16476, GSE62564 and GSE85047 datasets (Fig. 4a). Also, in E-MTAB-1781, GSE13136, GSE73517, TAR-GET, GSE16476, GSE62564 and GSE85047 datasets, the expression levels of ZNF436 were lower in neuroblastoma patients with age at diagnosis < 18months than neuroblastoma patients with age at diagnosis < 18months (Fig. 4b).

Moreover, contrast with stage 1 and stage 2 neuroblastoma, ZNF436 was down-regulated in stage 3 neuroblastoma patients in E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets (Fig. 4c). Also, contrast with stage 1 and stage 2 neuroblastoma, ZNF436 was down-regulated in stage 4 neuroblastoma patients in E-MTAB-1781, GSE13136, GSE73517, TARGET, GSE16476, GSE62564 and GSE85047 datasets (Fig. 4c). However, compared with stage 4, ZNF436 expressions in stage 4s neuroblastoma in E-MTAB-1781, GSE13136, GSE73517, TARGET, GSE16476, GSE62564 and GSE85047 datasets (Fig. 4c). However, compared with stage 4, ZNF436 expressions in stage 4s neuroblastoma in E-MTAB-1781, GSE13136, GSE73517, TARGET, GSE16476, GSE62564 and GSE85047 datasets were not significantly different (Fig. 4c).

ZNF436 is a predictor of the sub-types and overall survival of neuroblastoma

Our results suggested that ZNF436 was down-regulated with genetic 1p deletion or MYCN amplification and served as a favorable prognostic marker of neuroblastoma. Further, we attempted to determine the accuracy of ZNF436 in the prediction of 1p deletion, MYCN amplification and in the prediction of the overall survival of neuroblastoma. The ROC analysis in E-MTAB-1781 dataset indicated that ZNF436 could distinguish 1p deleted from 1p normal neuroblastoma patients with high specificity and sensitivity (Fig. 5a). Similar predictive specificity and sensitivity of ZNF436 in distinguishing MYCN amplified from MYCN non-amplified neuroblastoma patients was determined in E-MTAB-1781 dataset (Fig. 5a). Furthermore, ZNF436 also could predict the three years, five years or ten years overall survival of neuroblastoma with high accuracy in E-MTAB-1781 dataset (Fig. 5a).

The predictions of MYCN amplification and overall survival of neuroblastoma based on ZNF436 expression were validated in TARGET, GSE16476 and GSE62564 datasets. Similarly, ZNF436 had robust predictive values of MYCN amplification. In TARGET, GSE16476 and GSE62564 neuroblastoma cohorts, ROC curves showed the high specificity and sensitivity of ZNF436 in distinguishing MYCN amplified from MYCN non-amplified neuroblastoma patients (Fig. 5b). Furthermore, ZNF436 also could predict the three years, five years or ten years overall survival of neuroblastoma with high accuracy in GSE16476 and GSE62564 datasets (Fig. 5c). However, prediction of overall survival of neuroblastoma in TAR-GET dataset was not accurate (Fig. 5c). All those results highlighted the sub-types and overall survival predictions of ZNF436 in neuroblastoma.

ZNF436 is an Independent prognostic factor of neuroblastoma

The associations of age of diagnosis, MYCN amplification, 1p deletion and ZNF436 expression in the prediction of the overall survival of neuroblastoma were further analyzed using multivariate cox regression assay. Age of diagnosis, MYCN amplification and 1p deletion were independent prognostic factors of neuroblastoma, while, ZNF436 was not an independent prognostic factor of neuroblastoma in E-MTAB-1781 dataset (Fig. 6a). However, ZNF436 was a prognostic factor of neuroblastoma in E-MTAB-1781 dataset independent of age of diagnosis and MYCN amplification (Fig. 6b).

Furthermore, age of diagnosis was an independent prognostic factor in TARGET, GSE62564 and GSE85047 datasets, while not in GSE16476 dataset (Fig. 6c). MYCN amplification was also an independent prognostic factor in GSE16476, GSE62564 and GSE85047 datasets, but not in TARGET dataset (Fig. 6c). ZNF436 was an independent prognostic factor of neuroblastoma in TAR-GET, GSE16476 and GSE62564 neuroblastoma cohorts (Fig. 6c).

Synergistic prognostic effects of ZNF436 with MYCN amplification or age of diagnosis in neuroblastoma

Since, age of diagnosis, MYCN amplification and ZNF436 expression were independent prognostic factors in most neuroblastoma cohorts, we speculated the superior prognostic effects in the combinations of ZNF436 with MYCN amplification or age of diagnosis. Based on MYCN amplification and ZNF436 expression level, neuroblastoma patients were classified into different sub-groups. Neuroblastoma patients with ZNF436 high expressions and without MYCN amplification had significantly longer overall survival in E-MTAB-1781 and TAR-GET datasets (Fig. 7a). On the contrary, neuroblastoma patients with ZNF436 low expressions and with MYCN amplification had significantly shorter overall survival in E-MTAB-1781 and TARGET datasets (Fig. 7a). However, the synergistic prognostic effects of ZNF436 with MYCN amplification in GSE62564 dataset were not significant (Fig. 7a).

Neuroblastoma patients with age at diagnosis≥18months and with ZNF436 lower expressions had the worst prognosis in E-MTAB-1781, TARGET and GSE62564 datasets (Fig. 7b). On the contrary, neuroblastoma patients with age at diagnosis<18months and



Fig. 4 Expressions of ZNF436 in different sub-types of neuroblastoma. (a) Box plots showed the relative ZNF436 expression levels in neuroblastoma patients with or without MYCN amplification in E-MTAB-1781, GSE13136, GSE73517, TARGET, GSE16476, GSE62564 and GSE85047 datasets. P values were performed using Student's t test. (b) The relative ZNF436 expression levels in neuroblastoma patients with age of diagnosis \geq 18month or < 18months in E-MTAB-1781, GSE16476, GSE62564 and GSE85047 datasets. (c) The relative ZNF436 expression levels in neuroblastoma patients with different tumor stages in E-MTAB-1781, GSE13517, GSE16476, GSE62564 and GSE85047 datasets.



Fig. 5 ZNF436 is a predictor of the sub-types and overall survival of neuroblastoma. (**a**) The ROC curves showed the specificity and sensitivity of ZNF436 in the prediction of 1p deletion, MYCN amplification and in the prediction of the overall survival of neuroblastoma in E-MTAB-1781 dataset. (**b**) Prediction of MYCN amplification by ZNF436 in TARGET, GSE16476 and GSE62564 datasets. (**c**) Prediction of the overall survival of neuroblastoma by ZNF436 in TARGET, GSE16476 and GSE62564 datasets.



Fig. 6 ZNF436 is an independent prognostic factor of neuroblastoma. (a) Forest plots showed the associations of age, MYCN amplification, 1p deletion and ZNF436 with the overall survival of neuroblastoma in E-MTAB-1781 dataset. Hazard ratio and P values were determined by multivariate cox regression assay. (b) Forest plots showed the associations of age, MYCN amplification and ZNF436 with the clinical overall survival of neuroblastoma in E-MTAB-1781 dataset. (c) The associations of age, MYCN amplification and ZNF436 with the overall survival of neuroblastoma in TARGET, GSE16476, GSE62564 and GSE85047 datasets



Fig. 7 Synergistic prognostic effects of ZNF436 with MYCN amplification or age of diagnosis in neuroblastoma. (**a**) Neuroblastoma patients were classified into different sub-groups based on ZNF436 and MYCN amplification. Overall survival of each sub-group was determined using the Kaplan-Meier survival analysis in E-MTAB-1781, TARGET and GSE62564 datasets. P values were generated using log-rank test. (**b**) Neuroblastoma patients were classified into different sub-groups based on the ZNF436 and age of diagnosis. Overall survival of each sub-group was determined in E-MTAB-1781, TARGET and GSE62564 datasets

with ZNF43 higher expressions had the best prognosis in E-MTAB-1781 and TARGET datasets (Fig. 7b). Neuroblastoma patients with age at diagnosis < 18months and with ZNF43 higher expressions had the medium clinical survival in E-MTAB-1781, TARGET and GSE62564 datasets (Fig. 7b). Also, the synergistic prognostic effects of ZNF436 with age at diagnosis in GSE62564 dataset were not significant (Fig. 7b). Those results highlighted the complex of neuroblastoma cohorts from different datasets and some results from one dataset could not be validated by other datasets.

Construction of nomogram model to predict the overall survival of neuroblastoma

Our previous results suggested that ZNF436 was associated with the overall survival of neuroblastoma and showed the synergistic prognostic effects of ZNF436 with MYCN amplification or age of diagnosis in neuroblastoma. We then constructed a nomogram model based on age of diagnosis, MYCN amplification and ZNF436 to predict the overall survival of neuroblastoma in E-MTAB-1781, TARGET, GSE16476 and GSE62564 datasets (Fig. 8a). The accuracy of the nomogram model was further evaluated using calibration diagram. The C index values indicated the high accuracy of the nomogram models (Fig. 8b).

The risk point of each neuroblastoma patient in E-MTAB-1781, TARGET, GSE16476 and GSE62564 datasets was calculated from the nomogram models. Neuroblastoma with lower risk points had significantly longer overall survival than neuroblastoma with higher risk points in E-MTAB-1781, TARGET, GSE16476 and GSE62564 datasets (Fig. 9a). Moreover, in E-MTAB-1781, TARGET, GSE16476 and GSE62564

а	E-MTAB-1781	b	E-MTAB-1781
Points Age MYCN ZNF436 Total Points Linear Predicto 3-year Surviva 5-year Surviva	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Z Actual 5 vears OS (proportion)	C index: 0.84 C inde
Points Age MYCN ZNF436 Total Points Linear Predicto 3-year Surviva 5-year Surviva	TARGET 0 10 20 30 40 50 60 70 80 90 100 0 4 8 12 18 , Amp Not amp 8 7.8 7.4 7 6.8 6.4 6 5.8 5.4 0 10 20 30 40 50 60 70 80 90 110 130 -2 -1.5 -1 -0.5 0 0.5 1 1.5 2 0.9 0.8 0.7 0.6 0.4 0.2 0.1	Z Actual 5 vears OS (proportion)	TARGET C index: 0.65 C index: 0.65
Points Age MYCN ZNF436 Total Points Linear Predicto	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Actual 5 vears OS (proportion)	GSE16476 C index: 0.77

MYCN	Not arfip
ZNF436	170 150 130 110 90 70 50 30
Total Points	0 20 40 60 80 100 120 140 160 180 200
Linear Predictor	
3−year Survival	0.9 0.8 0.7 0.60.50.40.30.2 0.1
5−year Survival	0.9 0.7 0.5 0.3 0.1

	GSE62564
Points	0 <u>1020304050607080901</u> 00
Age	
MYCN	Not amp
ZNF436	
Total Points	0 20 40 60 80 100 120 140 160 180 200
Linear Predictor	
3-year Survival	
5−year Survival	0.9 0.8 0.7 0.5 0.3 0.1



0.4 0.6

Nomogram-Predicted Probability of 5 years OS

n=88 d=33 p=3, 39 subjects per group

0.8 1.0

0.0

0.2



Fig. 8 Construction of nomogram risk model based on age, MYCN amplification and ZNF436. (a) The nomogram models based on age, MYCN amplification and ZNF436 in TARGET, GSE16476, GSE62564 and GSE85047 datasets. (b) The calibration diagrams showed the accuracy of the nomogram models as evaluated using C index in TARGET, GSE16476, GSE62564 and GSE85047 datasets



Fig. 9 Predictive accuracy of the nomogram model. (a) Different overall survival of neuroblastoma with the low risk and high risk in TARGET, GSE16476, GSE62564 and GSE85047 datasets. (b) The ROC curves showed the predictive specificity and sensitivity of the risk points. AUC was calculated in the prediction of the overall survival of neuroblastoma in TARGET, GSE16476, GSE62564 and GSE85047 datasets

datasets, the risk points could predict the three years, five years or ten years overall survival of neuroblastoma with high specificity and sensitivity (Fig. 9b). More importantly, the predictive significance the nomogram model was higher than simple ZFN436 expression. For example, the five years overall survival prediction by ZFN436 expression was 0.76 (Fig. 5a), while, the five years overall survival prediction by the nomogram model was 0.87 in E-MTAB-1781 dataset (Fig. 9b).

Discussion

Genetic copy number variations, particularly MYCN amplification and 1p deletion, are implicated in the progress of neuroblastoma. The amplified or deleted DNA regions can be as large as 1 Mb and include multiple suspected neuroblastoma driver or suppressor genes [39, 40]. Determining the driver or suppressor roles of correlated genes could provide more prognostic makers and potential therapeutic targets for neuroblastoma. Previously, we had analyzed the MYCN regulated genes [41], E2F regulated genes [42] and TP53 regulated genes [43] for the prognosis of neuroblastoma. Using similar datasets, in this study, we analyzed the genetic loss of 1p, and investigated the expressions and prognosis of 1p candidate genes in neuroblastoma.

Contrast with gain of 2p24.3 MYCN regions, deletions of 1p regions influenced more genes. Based on TARGET dataset, 773 genes in chromosomal 1p regions were deleted in neuroblastoma. Some genes at this chromosomal regions are associated with low risks of neuroblastoma [26]. However, not all 1p candidate genes are down-regulated with the deletion of 1p. Compared with 1p normal neuroblastoma patients, only 105 genes in 1p regions were differentially expressed in 1p deleted neuroblastoma in E-MTAB-1781 dataset. And only 11 genes were commonly changed in 1p deleted neuroblastoma patients in E-MTAB-1781, GSE13136 and GSE73517 datasets. Only ZNF436 was involved in the prognosis of neuroblastoma in all E-MTAB-1781, TARGET, GSE16476, GSE62564 and GSE85047 datasets.

ZNF436 is a transcription factor belonging to the zinc finger protein family and modulates genes expressions through binding to the DNA elements [44]. The functions of ZNF436 in neuroblastoma are never reported. In glioma, ZNF436 could promote tumor cell proliferation [45]. In breast cancer, high ZNF436 expression is associated with high metastasis [46]. Our results provided new prognostic functions of ZNF436 in neuroblastoma and those prognostic functions may be associated with unique MYCN amplification and 1p deletion. ZNF436 is located in the chromosomal 1p36.12 regions. Consistent with the poor prognosis of 1p36 deletion, lower expression of ZNF436 was associated with worse clinical outcomes of neuroblastoma. Moreover, the expression levels of ZNF436 were lower in neuroblastoma with MYCN amplification or age at diagnosis \geq 18months, or in stage 4 neuroblastoma. ZNF436 had robust predictive values of MYCN amplification and overall survival of neuroblastoma. Furthermore, the prognostic significance of ZNF436 in neuroblastoma was independent of MYCN amplification and age of diagnosis. Combinations of ZNF436 expression with MYCN amplification or age of diagnosis achieved better prognosis of neuroblastoma.

Nomogram is extensively used in the prognosis of cancers [47–49] as well as obstetric diseases [50–53] and achieves accurate predictions. However, predictive nomogram models for the risks of neuroblastoma are unclear. So, in this study, using independent risk factors age of diagnosis, MYCN amplification and ZNF436, we developed a nomogram model to predict the overall survival of neuroblastoma. Compared with simple ZFN436 expression, the nomogram model could predict the overall survival of neuroblastoma with higher specificity and sensitivity.

To our best knowledge, this is the first integrated analysis of the expressions and prognosis of 1p candidate genes in neuroblastoma. Our results suggested that ZNF436 was served as prognostic biomarker of neuroblastoma. However, those results were generated using published neuroblastoma cohorts and lacked of additional in *vitro* and in *vivo* validations. Therefore, the detailed roles of ZNF436 should be further revealed using neuroblastoma cells and neuroblastoma patients.

Conclusions

ZNF436 was down-regulated in 1p deleted neuroblastoma and ZNF436 was lower in neuroblastoma patients with MYCN amplification or age at diagnosis \geq 18months, or with stage 4 neuroblastoma. Higher ZNF436 expression was associated with the longer event free survival and overall survival of neuroblastoma. ZNF436 had robust predictive values of MYCN amplification and overall survival of neuroblastoma. A nomogram risk model based on age, MYCN amplification and ZNF436 could predict the overall survival of neuroblastoma with higher specificity and sensitivity.

Abbreviations

TARGET	Therapeutically Applicable Research to Generate Effective
	Treatments
EMBL-EBI	The European Molecular Biology Laboratory-European
	Bioinformatics Institute
GEO	Gene Expression Omnibus
ZNF436	Zinc finger protein 436
HR	Hazard ratio
ROC	Receiver operating characteristic

AUC Area under the ROC curve

Acknowledgements

Not applicable.

Authors' contributions

HW designed the study and wrote the manuscript. HW, XW and LX performed the data analysis. JZ supervised the work.

Funding

The present study was sponsored by the Fujian Provincial Health Technology Project (grant no: 2021GGA049).

Data Availability

The datasets generated and/or analysed during the current study are available from the TARGET (https://ocg.cancer.gov/), EMBL-EBI (https://www.ebi.ac.uk/ arrayexpress/) and the GEO websites (www.ncbi.nlm.nih.gov/geo).

Declarations

Ethics approval and consent to participate Not applicable.

rior applicable

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 25 April 2023 / Accepted: 16 October 2023 Published online: 31 October 2023

References

- Maris JM, Hogarty MD, Bagatell R, Cohn SL. Neuroblastoma. Lancet. 2007;369(9579):2106–20.
- Maris JM. Recent advances in neuroblastoma. N Engl J Med. 2010;362(23):2202–11.
- Irwin MS, Park JR. Neuroblastoma: paradigm for precision medicine. Pediatr Clin North Am. 2015;62(1):225–56.
- 4. Davidoff AM. Neuroblastoma. Semin Pediatr Surg. 2012;21(1):2–14.
- Brodeur GM. Neuroblastoma: biological insights into a clinical enigma. Nat Rev Cancer. 2003;3(3):203–16.
- Cohn SL, Pearson AD, London WB, Monclair T, Ambros PF, Brodeur GM, Faldum A, Hero B, lehara T, Machin D, et al. The International Neuroblastoma Risk Group (INRG) classification system: an INRG Task Force report. J Clin Oncol. 2009;27(2):289–97.
- Defferrari R, Tonini GP, Conte M, Papio F, Sementa AR, Valent A, Schena F, Perri P, Mazzocco K. Concomitant DDX1 and MYCN gain in neuroblastoma. Cancer Lett. 2007;256(1):56–63.
- Weber A, Imisch P, Bergmann E, Christiansen H. Coamplification of DDX1 correlates with an improved survival probability in children with MYCNamplified human neuroblastoma. J Clin Oncol. 2004;22(13):2681–90.
- Ackermann S, Cartolano M, Hero B, Welte A, Kahlert Y, Roderwieser A, Bartenhagen C, Walter E, Gecht J, Kerschke L, et al. A mechanistic classification of clinical phenotypes in neuroblastoma. Science. 2018;362(6419):1165–70.
- Attiyeh EF, London WB, Mosse YP, Wang Q, Winter C, Khazi D, McGrady PW, Seeger RC, Look AT, Shimada H, et al. Chromosome 1p and 11q deletions and outcome in neuroblastoma. N Engl J Med. 2005;353(21):2243–53.
- 11. Spitz R, Hero B, Simon T, Berthold F. Loss in chromosome 11q identifies tumors with increased risk for metastatic relapses in localized and 4S neuroblastoma. Clin Cancer Res. 2006;12(11 Pt 1):3368–73.
- Lastowska M, Cotterill S, Pearson AD, Roberts P, McGuckin A, Lewis I, Bown N. Gain of chromosome arm 17q predicts unfavourable outcome in neuroblastoma patients. U.K. children's Cancer Study Group and the U.K. Cancer Cytogenetics Group. Eur J Cancer. 1997;33(10):1627–33.
- Caron H, van Sluis P, de Kraker J, Bokkerink J, Egeler M, Laureys G, Slater R, Westerveld A, Voute PA, Versteeg R. Allelic loss of chromosome 1p as a

predictor of unfavorable outcome in patients with neuroblastoma. N Engl J Med. 1996;334(4):225–30.

- White PS, Thompson PM, Gotoh T, Okawa ER, Igarashi J, Kok M, Winter C, Gregory SG, Hogarty MD, Maris JM, et al. Definition and characterization of a region of 1p36.3 consistently deleted in neuroblastoma. Oncogene. 2005;24(16):2684–94.
- Kuick CH, Tan JY, Jasmine D, Sumanty T, Ng AYJ, Venkatesh B, Chen H, Loh E, Jain S, Seow WY, et al. Mutations of 1p genes do not consistently abrogate Tumor suppressor functions in 1p-intact neuroblastoma. BMC Cancer. 2022;22(1):717.
- Fransson S, Martinsson T, Ejeskar K. Neuroblastoma tumors with favorable and unfavorable outcomes: significant differences in mRNA expression of genes mapped at 1p36.2. Genes Chromosomes Cancer. 2007;46(1):45–52.
- 17. Thompson PM, Gotoh T, Kok M, White PS, Brodeur GM. CHD5, a new member of the chromodomain gene family, is preferentially expressed in the nervous system. Oncogene. 2003;22(7):1002–11.
- Okawa ER, Gotoh T, Manne J, Igarashi J, Fujita T, Silverman KA, Xhao H, Mosse YP, White PS, Brodeur GM. Expression and sequence analysis of candidates for the 1p36.31 Tumor suppressor gene deleted in neuroblastomas. Oncogene. 2008;27(6):803–10.
- Koyama H, Zhuang T, Light JE, Kolla V, Higashi M, McGrady PW, London WB, Brodeur GM. Mechanisms of CHD5 inactivation in neuroblastomas. Clin Cancer Res. 2012;18(6):1588–97.
- Garcia I, Mayol G, Rodriguez E, Sunol M, Gershon TR, Rios J, Cheung NK, Kieran MW, George RE, Perez-Atayde AR, et al. Expression of the neuron-specific protein CHD5 is an Independent marker of outcome in neuroblastoma. Mol Cancer. 2010;9:277.
- Laut AK, Dorneburg C, Furstberger A, Barth TFE, Kestler HA, Debatin KM, Beltinger C. CHD5 inhibits Metastasis of neuroblastoma. Oncogene. 2022;41(5):622–33.
- Romani M, Tonini GP, Banelli B, Allemanni G, Mazzocco K, Scaruffi P, Boni L, Ponzoni M, Pagnan G, Raffaghello L, et al. Biological and clinical role of p73 in neuroblastoma. Cancer Lett. 2003;197(1–2):111–7.
- 23. Horvilleur E, Bauer M, Goldschneider D, Mergui X, de la Motte A, Benard J, Douc-Rasy S, Cappellen D. p73alpha isoforms drive opposite transcriptional and post-transcriptional regulation of MYCN expression in neuroblastoma cells. Nucleic Acids Res. 2008;36(13):4222–32.
- De Laurenzi V, Raschella G, Barcaroli D, Annicchiarico-Petruzzelli M, Ranalli M, Catani MV, Tanno B, Costanzo A, Levrero M, Melino G. Induction of neuronal differentiation by p73 in a neuroblastoma cell line. J Biol Chem. 2000;275(20):15226–31.
- Chen ZX, Wallis K, Fell SM, Sobrado VR, Hemmer MC, Ramskold D, Hellman U, Sandberg R, Kenchappa RS, Martinson T, et al. RNA helicase A is a downstream mediator of KIF18beta tumor-suppressor function in neuroblastoma. Cancer Discov. 2014;4(4):434–51.
- Yeh IT, Lenci RE, Qin Y, Buddavarapu K, Ligon AH, Leteurtre E, Do Cao C, Cardot-Bauters C, Pigny P, Dahia PL. A germline mutation of the KIF1B beta gene on 1p36 in a family with neural and nonneural tumors. Hum Genet. 2008;124(3):279–85.
- 27. Ma X, Liu Y, Liu Y, Alexandrov LB, Edmonson MN, Gawad C, Zhou X, Li Y, Rusch MC, Easton J, et al. Pan-cancer genome and transcriptome analyses of 1,699 paediatric leukaemias and solid tumours. Nature. 2018;555(7696):371–6.
- Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal. 2013;6(269):pl1.
- Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, Jacobsen A, Byrne CJ, Heuer ML, Larsson E, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov. 2012;2(5):401–4.
- Oberthuer A, Juraeva D, Hero B, Volland R, Sterz C, Schmidt R, Faldum A, Kahlert Y, Engesser A, Asgharzadeh S, et al. Revised risk estimation and treatment stratification of low- and intermediate-risk neuroblastoma patients by integrating clinical and molecular prognostic markers. Clin Cancer Res. 2015;21(8):1904–15.
- Lastowska M, Viprey V, Santibanez-Koref M, Wappler I, Peters H, Cullinane C, Roberts P, Hall AG, Tweddle DA, Pearson AD, et al. Identification of candidate genes involved in neuroblastoma progression by combining genomic and expression microarrays with survival data. Oncogene. 2007;26(53):7432–44.
- 32. Henrich KO, Bender S, Saadati M, Dreidax D, Gartlgruber M, Shao C, Herrmann C, Wiesenfarth M, Parzonka M, Wehrmann L, et al. Integrative Genome-Scale

Analysis Identifies Epigenetic Mechanisms of Transcriptional Deregulation in unfavorable neuroblastomas. Cancer Res. 2016;76(18):5523–37.

- Molenaar JJ, Koster J, Zwijnenburg DA, van Sluis P, Valentijn LJ, van der Ploeg I, Hamdi M, van Nes J, Westerman BA, van Arkel J, et al. Sequencing of neuroblastoma identifies chromothripsis and defects in neuritogenesis genes. Nature. 2012;483(7391):589–93.
- Molenaar JJ, Domingo-Fernandez R, Ebus ME, Lindner S, Koster J, Drabek K, Mestdagh P, van Sluis P, Valentijn LJ, van Nes J, et al. LIN28B induces neuroblastoma and enhances MYCN levels via let-7 suppression. Nat Genet. 2012;44(11):1199–206.
- Lamers F, Schild L, Koster J, Speleman F, Ora I, Westerhout EM, van Sluis P, Versteeg R, Caron HN, Molenaar JJ. Identification of BIRC6 as a novel intervention target for neuroblastoma therapy. BMC Cancer. 2012;12:285.
- Wang C, Gong B, Bushel PR, Thierry-Mieg J, Thierry-Mieg D, Xu J, Fang H, Hong H, Shen J, Su Z, et al. The concordance between RNA-seq and microarray data depends on chemical treatment and transcript abundance. Nat Biotechnol. 2014;32(9):926–32.
- Rajbhandari P, Lopez G, Capdevila C, Salvatori B, Yu J, Rodriguez-Barrueco R, Martinez D, Yarmarkovich M, Weichert-Leahey N, Abraham BJ, et al. Cross-cohort Analysis identifies a TEAD4-MYCN positive feedback Loop as the Core Regulatory element of high-risk neuroblastoma. Cancer Discov. 2018;8(5):582–99.
- Campbell K, Gastier-Foster JM, Mann M, Naranjo AH, Van Ryn C, Bagatell R, Matthay KK, London WB, Irwin MS, Shimada H, et al. Association of MYCN copy number with clinical features, Tumor biology, and outcomes in neuroblastoma: a report from the Children's Oncology Group. Cancer. 2017;123(21):4224–35.
- 39. Kao J, Pollack JR. RNA interference-based functional dissection of the 17q12 amplicon in Breast cancer reveals contribution of coamplified genes. Genes Chromosomes Cancer. 2006;45(8):761–9.
- Scott D, Elsden J, Pearson A, Lunec J. Genes co-amplified with MYCN in neuroblastoma: silent passengers or co-determinants of phenotype? Cancer Lett. 2003;197(1–2):81–6.
- Wang H, Wang X, Xu L, Zhang J, Cao H. Prognostic significance of MYCN related genes in neuroblastoma: a study based on TARGET and GEO datasets. BMC Pediatr. 2020;20(1):314.
- Wang H, Wang X, Xu L, Zhang J. Prognostic analysis of E2F transcription factors E2F1 and E2F3 in four Independent neuroblastoma cohorts. BMC Pediatr. 2022;22(1):376.
- Wang H, Wang X, Xu L, Zhang J. TP53 and TP53-associated genes are correlated with the prognosis of paediatric neuroblastoma. BMC Genom Data. 2022;23(1):41.
- Razin SV, Borunova VV, Maksimenko OG, Kantidze OL. Cys2His2 zinc finger protein family: classification, functions, and major members. Biochem (Mosc). 2012;77(3):217–26.
- 45. Shang Y, Li Y, Zhang Y, Wang J. ZNF436 promotes Tumor cell proliferation through transcriptional activation of BCL10 in glioma. Biochem Biophys Res Commun. 2019;515(4):572–8.
- 46. Chen Z, Cui N, Zhao JS, Wu JF, Ma F, Li C, Liu XY. Expressions of ZNF436, betacatenin, EGFR, and CMTM5 in Breast cancer and their clinical significances. Eur J Histochem 2021, 65(1).
- Liang W, Zhang L, Jiang G, Wang Q, Liu L, Liu D, Wang Z, Zhu Z, Deng Q, Xiong X, et al. Development and validation of a nomogram for predicting survival in patients with resected non-small-cell Lung cancer. J Clin Oncol. 2015;33(8):861–9.
- Su J, Miao LF, Ye XH, Cui MS, He XF. Development of prognostic signature and nomogram for patients with Breast cancer. Med (Baltim). 2019;98(11):e14617.
- Wu J, Zhang H, Li L, Hu M, Chen L, Xu B, Song Q. A nomogram for predicting overall survival in patients with low-grade endometrial stromal sarcoma: a population-based analysis. Cancer Commun (Lond). 2020;40(7):301–12.
- Lv B, Zhang Y, Yuan G, Gu R, Wang J, Zou Y, Wei L. Establishment of a nomogram model for predicting adverse outcomes in advanced-age pregnant women with preterm preeclampsia. BMC Pregnancy Childbirth. 2022;22(1):221.
- Du J, Zhang X, Chai S, Zhao X, Sun J, Yuan N, Yu X, Zhang Q. Nomogrambased risk prediction of macrosomia: a case-control study. BMC Pregnancy Childbirth. 2022;22(1):392.
- 52. Zheng J, Zhang L, Zhou Y, Xu L, Zhang Z, Luo Y. Development and evaluation of a nomogram for adverse outcomes of preeclampsia in Chinese pregnant women. BMC Pregnancy Childbirth. 2022;22(1):504.

 Zou Y, Zhang Y, Yin Z, Wei L, Lv B, Wu Y. Establishment of a nomogram model to predict macrosomia in pregnant women with gestational Diabetes Mellitus. BMC Pregnancy Childbirth. 2021;21(1):581.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.