

CASE REPORT

Open Access

# Severe congenital neutropenia due to G6PC3 deficiency: early and delayed phenotype in two patients with two novel mutations

Lucia Dora Notarangelo<sup>1\*</sup>, Gianfranco Savoldi<sup>2</sup>, Sara Cavagnini<sup>1</sup>, Veronica Bennato<sup>1</sup>, Sabrina Vasile<sup>3</sup>, Alba Pilotta<sup>4</sup>, Alessandro Plebani<sup>5</sup> and Fulvio Porta<sup>1</sup>

## Abstract

Severe Congenital Neutropenia type 4 (SCN4, OMIM 612541) is a rare autosomal recessive disease due to mutations in the *G6PC3* gene. The phenotype comprises neutropenia of variable severity and other anomalies including congenital heart defects, prominent superficial veins, uro-genital anomalies, facial dysmorphism, growth and developmental delay and intermittent thrombocytopenia. In some patients, SCN represents the only manifestation of the disease. Variable findings have been reported at bone marrow examination ranging from a maturation arrest at the myelocyte/promyelocyte stage (either in a hypocellular or hypercellular context) to myelokathexis. Here we report two patients harbouring two novel mutations in the *G6PC3* gene, including the first Italian patient even described. Both the patients share profound neutropenia with severe infections early in life; in one case non-hematopoietic stigmata of the syndrome, including evident facial dysmorphism and vascular anomalies, appeared gradually over time, prominently in the second decade. Therefore, *G6PC3* defects should be considered in any case of congenital, unexplained neutropenia regardless of the clinical phenotype. Both patients are on G-CSF treatment with no evidence of malignant evolution. Even if *G6PC3* deficiency seems not to have a propensity towards malignancy, a careful evaluation is warranted.

**Keywords:** Neutropenia, Severe congenital neutropenia type 4, SCN4, *G6PC3*, G-CSF

## Background

Severe congenital neutropenia (SCN) is a heterogeneous group of mature neutrophil deficiency disorders with or without immunological or extra-hematopoietic abnormalities [1]. Various genetic defects can cause SCN, and some of them also predispose to MDS/AML [2]. SCN type 4 (SCN4, OMIM 612541) is a rare autosomal recessive disease due to mutations in the *G6PC3* gene which encodes for the catalytic subunit of the glucose-6-phosphatase 3 protein (G6Pase3) [3]. Along with G-6Pase, which is mutated in Glycogen Storage Disease Type 1 (GSD1), G-6Pase 3 belongs to the glucose-6-phosphatase (Glc-6-Pase) family, a group of transmembrane, endoplasmic reticulum (ER)- associated proteins [3]. Although the two isoenzymes share the same catalytic properties, only

defects in G-6Pase are associated with metabolic symptoms, reflecting their unique role in fasting glucose homeostasis and different tissue expression patterns [3].

The human *G6PC3* gene is located at chromosome 17q21 and has 6 exons. SNC4 is most often due to missense mutations. Moreover, some mutations occur with higher frequency in certain ethnic groups, possibly reflecting a founder effect [4,5]. Since the first description, 59 patients with SCN4 have been reported. All of them share neutropenia of variable severity and some have other anomalies including congenital heart defects, prominent superficial veins, uro-genital anomalies, facial dysmorphism, growth and developmental delay and intermittent thrombocytopenia [4,6]. In some patients, SCN represents the only manifestation of the disease [7]. Mutation of the same gene may give rise to Dursun Syndrome (DS) (OMIM 613034), a more severe, often lethal, phenotype [8]. This syndrome is characterized

\* Correspondence: [luciadora.notarangelo@spedalicivili.brescia.it](mailto:luciadora.notarangelo@spedalicivili.brescia.it)

<sup>1</sup>Pediatric Onco-haematology and BMT Unit, Children's Hospital, Spedali Civili of Brescia, Brescia, Italy

Full list of author information is available at the end of the article

by a triad of familial primary pulmonary hypertension (PPH), leucopenia and atrial septal defects [9]. Other features include secundum-type atrial septal defect, intermittent neutropenia, lymphopenia, monocytosis, anemia, and thymic hypoplasia. The prognosis is often poor due to severe respiratory distress [9]. Here we report two patients harbouring two novel mutations in *G6PC3* gene, including the first Italian patient who is also, to our knowledge, the longest survivor with *SCN4* on treatment with G-CSF ever described.

## Case report

### Patient 1

The proband is a 20 year old female, second child of healthy and unrelated Italian parents, born at term after an uneventful pregnancy. She developed recurrent and serious *S. aureus* infections since early in life (skin infection at 10 days of age, pulmonary abscess at 2 months, brain abscess leading to lateralized seizures at 5 months) (Table 1). Complete blood cell count showed normal white blood cell count (6.600/ $\mu$ L) with severe neutropenia

**Table 1 Hematological and immunological evaluation, phenotype and mutations in *G6PC3* patients**

	Patient		
	1	2	
Eth	Italian	Turkish (consanguinous)	
Sex	F	M	
Current age (years)	20	3	
Infections prior to G-CSF therapy	Skin, pulmonary and brain abscess by <i>S. Aureus</i>	Otitis, parotitis, <i>S. Viridans</i> sepsis, <i>S. Aureus</i> gluteal abscess, recurrent aphthous stomatitis	
Blood cells count	White blood cells ^ (4.5-17)	6,6	2,7
	Neutrophils ^ (1.5-6.0 $\times$ 109/L)	0,22	0,06
	Lymphocytes ^ (1.3-8.5 $\times$ 109/L)	2,9	1,9
	Monocytes ^ (0.1-1.0 $\times$ 109/L)	3,4	0,7
	Platelet count ^ (150-450 cells/ $\mu$ L)	610	347
	Intermittent thrombocytopenia	Yes (range 46-158 cells/ $\mu$ L)	No
Serum Immunoglobulin concentrations	IgG (mg/dL)	1.240 (231-947)	974 (462-1710)
	IgA (mg/dl)	54 (8-74)	64 (27-173)
	IgM (mg/dl)	79 (26-210)	118 (62-257)
	IgE (kU/L) (n.v. < 95)	3	7
Lymphocyte subset (cells/ $\mu$ L)	CD3+ (770-1880)	720	1287
	CD3 + CD4+ ( 470-1240)	373	805
	CD3 + CD8+ (215-730)	318	426
	CD19+ (100-390)	218	460
	CD16+ (70-550)	69	79
	CD11b (PMN)	98%	N.D.
	CD18+ (PMN)	100%	N.D.
	Extra haematological features	Facial dysmorphisms, mitral valve prolapse, prominent veins, ligamentous laxity, inguinal hernia, delayed puberty, hypergonadotrophic hypogonadism	Facial dysmorphisms, prominent veins, sensorineural hearing loss, micropenis, coronal ipospadia.
G6PC3	c.144C>A/c.373_375delAAT	c.680_684delinsT/ c.680_684delinsT	
Mutation	p.Y48*/ p.I125del	p.S227Lfs*3/ p.S227Lfs*3	

^ prior to G-CSF therapy.

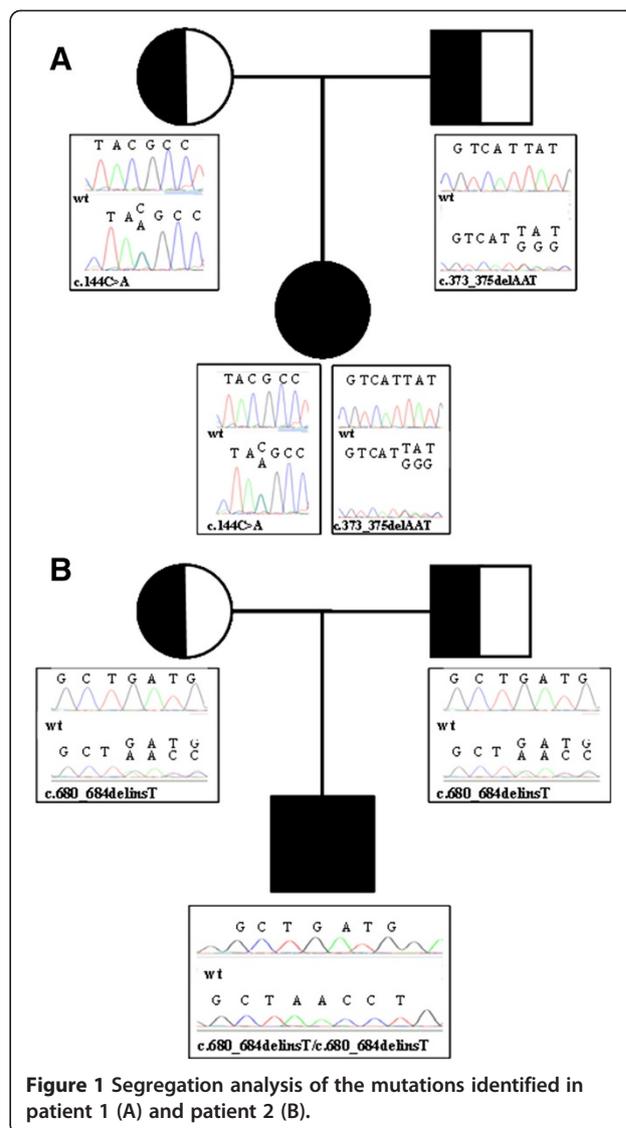
NA: not available.

Immunoglobulins normal value for age are reported in brackets.

(220 cells/ $\mu$ L), monocytosis (3,400 cells/ $\mu$ L), mild anemia (Hb 8.6 g/dl, MCV 78 fl) and thrombocytosis ( $610 \times 10^9/L$ ). Immunological evaluation, including serum immunoglobulins, lymphocyte subsets, expression of adhesion molecules, complement, dihydrorhodamine (DHR) test, was normal (Table 1). Due to persistent neutropenia, a bone marrow aspiration was performed, which showed global hypercellularity with myeloid hyperplasia, maturation arrest at the promyelocyte-myelocyte stage and paucity of mature neutrophils. Treatment with G-CSF was started (5–10  $\mu$ g/Kg/d) with normalization of ANC ( $>1,500$  cell/ $\mu$ l). At 8 yrs of age, the patient developed thrombocytopenia ( $33 \times 10^9/L$ ), which was initially attributed to G-CSF treatment. Subsequently the platelet count spontaneously increased with an intermittent course (Table 1). Reduced height growth speed was observed since 10 years of age. Because of delayed puberty, a decapeptyl test was performed at 16 years, and results were consistent with hypergonadotrophic hypogonadism (data not shown). A facial dysmorphism with frontal bossing, upturned nose and retrognathia became more evident over time, prominently in the second decade, along with the appearance of edema and superficial venous pattern of the lower limbs around 13 years. Other non hematological aspects are reported in Table 1. Upon informed consent, the coding region of the *G6PC3* gene was sequenced, revealing compound heterozygosity for c.144C > A substitution (predicted to result in p.Y48\* premature termination) and a novel three nucleotide deletion (c.373\_375delAAT), predicted to cause deletion of the Isoleucine residue at position 125 (p.I125del). Each parent was found to be heterozygous for one of the two mutations (see Figure 1A). In order to define the pathogenic role of the variants identified in our patients, we performed DHPLC analysis on 150 samples of genomic DNA (300 chromosomes) from unrelated healthy control individuals [10]. Analysis by DHPLC of the PCR fragments containing the mutations did not show any alterations of the DHPLC profiles in the control subjects analysed. The patient is currently 20 years old without major infections or developmental delay, on G-CSF treatment (2  $\mu$ g/Kg/d). Yearly bone marrow evaluations have confirmed previous findings.

#### Patient 2

A male patient, third child of consanguineous Turkish parents, was referred to us at the age of 2 years for severe neutropenia and early-onset recurrent infections including otitis, parotitis, *S. viridans* sepsis, *S. aureus* gluteal abscess, and recurrent aphthous stomatitis (Table 1). Mild sensorineural hearing loss was diagnosed at 4 months of life. At 6 months of age, the patient developed chickenpox without complications. Bone marrow aspiration, performed at another institution, revealed delayed granulocyte maturation.



**Figure 1** Segregation analysis of the mutations identified in patient 1 (A) and patient 2 (B).

No mutations were found in the *ELANE* and *HAX1* genes. On physical examination, the weight was 13 Kg (25-50<sup>th</sup> percentile), and length was 88 cm (50<sup>th</sup> percentile). There was facial dysmorphism with frontal bossing, upturned nose and malar hypoplasia, prominent superficial venous pattern on the arms, legs and trunk, and micropenis with coronal ipsipadia. Laboratory investigations revealed severe neutropenia (ANC: 140 cells/ $\mu$ L) with normal lymphocyte, monocyte and platelet counts, slight microcytic anemia (Hb 10,5 g/dl; MCV 65,4 fl), normal serum immunoglobulins, protective antibody response to tetanus toxoid, and normal distribution of lymphocyte subsets. Abdominal ultrasound and echocardiography were also normal (see Table 1). Upon informed consent, mutation analysis of the *G6PC3* gene was performed and revealed a novel homozygous 5 bp indel mutation (c.680\_684delinsT) in exon 6, resulting in Serine to Leucine substitution at position 227, and frameshift with

premature termination (p.S227Lfs\*3). Both parents were found heterozygous for the mutation (see Figure 1B). DHPLC analysis on 150 samples of genomic DNA (300 chromosomes) from unrelated healthy control individuals didn't show any alterations [10]. Treatment with G-CSF (5 µg/Kg/d) was started with rapid increase of ANC (12,200 cells/µL). Due to the appearance of thrombocytopenia ( $71 \times 10^9/L$ ) and splenomegaly, treatment with G-CSF was suspended with clinical and laboratory resolution and then re-started at the dose of 4 µg/Kg 3 times/week. At present, the patient is 3 years old, infection-free and his ANC is stably around 2,000 cells/µL. Other than mild and transient splenomegaly, no adverse events have been observed.

## Discussion

Biallelic mutations in the *G6PC3* gene are associated with a variable phenotype. In the two largest series reported until now [4,11] severe neutropenia predisposing to severe infections is the most reliable marker of disease, since congenital heart defects (mostly atrial septal defects) are present in around 80% of patients while prominent superficial veins and facial dysmorphism are frequent but less consistent. Furthermore, six patients with *G6PC3* mutations associated with non syndromic SCN have been described [7]. The role of *G6PC3* mutations in causing non hematological manifestations of SCN4 is not yet clear. Neutrophils and skin fibroblasts from SCN4 patients have an increased susceptibility to apoptosis through a mechanism that involves glycogen synthetase kinase 3 beta (GSK-3beta) and endoplasmic reticulum stress. The same mechanism also may play a role in causing dysmorphism [12]. Variable findings have been reported at bone marrow examination: most patients present a maturation arrest at the myelocyte/promyelocyte stage (either in a hypocellular or hypercellular context), however myelokathexis has been also observed [13]. No genotype-phenotype correlation has been demonstrated to explain the bone marrow variability as the same mutation can cause either maturation arrest or normo/hypercellular bone marrow [14]. On the contrary, the extra-hematological aspects of the syndrome could be possibly related to the variation in the residual activity of the enzyme, as well as to the genetic or environmental background, since specific missense mutations has been found in association with non-syndromic patients [4,7]. It has been postulated that missense mutations with a higher residual activity could predispose to a non -syndromic phenotype [7]. We herein report on two new cases of SCN4. In addition to contributing two novel *G6PC3* mutations, the patients described also offer some diagnostic and therapeutic observations. Patient 1 illustrates that non-hematopoietic stigmata of the syndrome (including evident facial dysmorphism

and vascular anomalies) may appear gradually over time, making the diagnosis difficult early in life. The six patients with non syndromic SCN4 reported were all in the first and second decade of life, the oldest one being 18 years old [7]. Longer follow up studies are needed to assess whether SCN4 is uniformly associated with extra-hematopoietic manifestations (which may also occur later in life) or whether in some patients the clinical phenotype may be restricted to SCN for the entire duration of life. Therefore, *G6PC3* mutation analysis should be considered in all cases of congenital, unexplained neutropenia, regardless of the presence of extra-hematopoietic manifestations or of parental consanguinity, once ELANE and HAX1 mutations are ruled out.

Growth retardation and delayed puberty have been reported in some patients with SCN4 [15]. In one case, hypogonadotropic hypogonadism was documented, and hypothyroidism was considered an extra-hematological manifestation of SCN4 in another patient [15,16]. Recently, testicular failure with very high levels of luteinizing hormone (LH) and follicle-stimulating hormone (FSH) has been reported in a *G6PC3* subject [17]. Patient 1 in this report showed delayed puberty with appearance of irregular menses at 17 years of life. Results of the decapeptyl test were consistent with hypergonadotropic hypogonadism, and additional tests demonstrated normal estrogen levels and increased LH values, as in partial ovarian insufficiency. The observation of hormonal and growth abnormalities in several patients suggest that appropriate endocrine evaluation should be considered in SCN4 patients.

Sensorineural hearing loss has been previously reported in five patients, all of which were homozygous for a *G6PC3* mutation; moreover parental consanguinity was documented or postulated in several of them [4]. Patient 2 in this report also suffered from sensorineural deafness. Given the relatively high frequency of genetically determined sensorineural deafness in the general population, it remains to be determined whether the latter represents a novel syndromic aspect of the disease, or whether parental consanguinity may have caused simultaneous inheritance of distinct, autosomal recessive genetic defects in *G6PC3* and in another gene responsible for sensorineural deafness.

The *G6PC3* protein is a paralog of *G6PC* with which it shares 36% sequence identity [3]. Since the *G6PC3* protein structure is not well defined, the functional consequences of the mutations are predicted based on studies performed on the better known *G6PC* protein. The I125del mutation in patient 1 is located in the transmembrane region and may interfere with the correct folding and stability, in analogy with the effect of missense, transmembrane mutations in the *G6PC* protein [4]. Patient 2 presents a new complex mutation in exon

6 resulting in amino acid substitution in the endoluminal domain with a premature stop codon after three amino acids. Although no functional studies have been performed to investigate the consequences of this mutation, the occurrence of a premature termination likely interferes with both the structure and the function of the protein.

Recombinant human G-CSF has been successfully employed in the treatment of congenital neutropenia since the end of 80's, with reduction of the rate of severe infections and improved quality of life, even if some short and long-term adverse effects have been reported [18,19]. Thrombocytopenia and splenomegaly are frequent complications of G-CSF treatment. In patient 1 thrombocytopenia was at first attributed to G-CSF therapy; however, its intermittent course, not related to changes of G-CSF dosage, is similar to what is reported in 10% of *G6PC3* patients, and is therefore most likely related to the disease itself [11]. In patient 2, both splenomegaly and thrombocytopenia were possibly due to the G-CSF treatment, since they regressed after treatment discontinuation, with subsequent reappearance of a mild splenomegaly even at lower doses of G-CSF.

Leukemic transformation is well documented in patients with SCN on G-CSF treatment. Factors contributing to leukemic transformation include the genetic type and severity of neutropenia and G-CSF exposure, i.e. cumulative dose as well as mean dose per injection [20]. Interestingly, no cases of leukemia have been reported in patients with SCN4, possibly due to the young age and the small size of the cohort described [11]. Patient 1 in this report has been treated with G-CSF continuously for 20 years, at a median dose of 3.9 µg/Kg/day (range 2.2-5), with no evidence of leukemia or myelodysplasia on repeated examinations of bone marrow specimens. Nevertheless, a careful evaluation of a possible malignant evolution is warranted.

## Consent

Written informed consent was obtained from the patients for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

## Abbreviations

SCN4: Severe Congenital Neutropenia type 4; *G6PC3*: glucose 6 phosphatase catalytic subunit 3; G-CSF: Granulocyte colony-stimulating factor; MDS/AML: Myelodysplasia/acute myeloid leukemia; *G6Pase3*: Glucose-6-phosphatase 3 protein; GSD1: Glycogen Storage Disease Type 1; DS: Dursun syndrome; PPH: Primary pulmonary hypertension; DHR: Dihydrorhodamine; ANC: Absolute neutrophil count; DHPLC: Denaturing High Pressure Liquid Chromatography; GSK-3beta: Glycogen synthetase kinase 3 beta; LH: Luteinizing hormone; FSH: Follicle-stimulating hormone.

## Competing interest

The authors declare that they have no competing interest.

## Authors' contributions

LDN, SC, VB, SB, API and FP contributed to the diagnosis and provided clinical assistance; GS performed the molecular analysis; API performed the endocrine investigation, LDN reviewed the literature, collected the data and wrote the paper; API and GS revised critically the manuscript. All authors read and approved the final version.

## Acknowledgements

We thank ABE (Associazione Bambino Emopatico, Brescia) for its contribution.

## Author details

<sup>1</sup>Pediatric Onco-haematology and BMT Unit, Children's Hospital, Spedali Civili of Brescia, Brescia, Italy. <sup>2</sup>Department of Pathology, Laboratory of Genetic Disorders of Childhood, A. Nocivelli Institute for Molecular Medicine, Spedali Civili, Brescia, Italy. <sup>3</sup>U.O. di Pediatria, Ospedale Valduce, Como, Italy. <sup>4</sup>Pediatrics Clinic, Spedali Civili of Brescia, Brescia, Italy. <sup>5</sup>Department of Clinical and Experimental Science, Pediatrics Clinic and Institute of Molecular Medicine "A. Nocivelli", University of Brescia and Spedali Civili of Brescia, Brescia, Italy.

Received: 25 June 2014 Accepted: 10 October 2014

Published online: 14 November 2014

## References

1. Donadieu J, Fenneteau O, Beaupain B, Mahlaoui N, Bellanné Chantelot C: **Congenital neutropenia: diagnosis, molecular bases and patient management.** *Orphanet J Rare Dis* 2011, **6**:26.
2. Botzug K, Klein C: **Genetic etiologies of severe congenital neutropenia.** *Curr Opin Pediatr* 2011, **23**(1):21–26.
3. Ghosh A, Shieh JJ, Pan CJ, Yang Chou J: **Histidine 167 Is the Phosphate Acceptor in Glucose-6-phosphate-β Forming a Phosphohistidine Enzyme Intermediate during Catalysis.** *J Biol Chem* 2004, **279**:12479–12483.
4. Banka S, Newman WG: **A clinical and molecular review of ubiquitous glucose-6 phosphatase deficiency caused by *G6PC3* mutations.** *Orphanet J Rare Dis* 2013, **8**:84.
5. Bradley NS, Evans C, Ali A, Ancliff PJ, Hayee B, Segal AW, Hall G, Kaya Z, Shakoori AR, Linch DC, Gale RE: **Phenotypic heterogeneity and evidence of a founder effect associated with *G6PC3* mutations in patients with severe congenital neutropenia.** *Br J Haematol* 2012, **158**:138–152.
6. Botzug K, Appaswamy G, Ashikov A, Schaffer AA, Salzer U, Diestelhorst J, Germeshausen M, Brandes G, Lee-Gossler J, Noyan F, Gatzke AK, Minkov M, Greil J, Kratz C, Petropoulou T, Pellier I, Bellanné-Chantelot C, Rezaei N, Monkemoller K, Irani-Hakimeh N, Hans Bakker H, Gerardy-Schahn R, Zeidler C, Grimbacher B, Welte K, Klein C: **A syndrome with Congenital Neutropenia and Mutations in *G6PC3*.** *N Engl J Med* 2009, **360**:32–43.
7. Banka S, Wynn R, Byers H, Arkwright PD, Newman WG: ***G6PC3* mutations cause non-syndromic severe congenital neutropenia.** *Mol Genet Metab* 2013, **108**:138–141.
8. Banka S, Newman WG, Ozgul RK, Dursun A: **Mutations in the *G6PC3* gene cause Dursun syndrome.** *Am J Med Genet* 2010, **152**:2609–2611.
9. Dursun A, Ozgul RK, Soydas A, Tugrul T, Gurgey A, Celiker A, Barst RJ, Knowles JA, Mahesh M, Morse JH: **Familial pulmonary arterial hypertension, leucopenia, and atrial septal defect: A probable new familial syndrome with multisystem involvement.** *Clin Dysmorphol* 2009, **18**:19–23.
10. Fackenthal DL, Chen PX, Howe T, Das S: **Denaturing high-performance liquid chromatography for mutation detection and genotyping.** *Methods Mol Biol* 2013, **1015**:25–54.
11. Botzug K, Rosenberg PS, Dorda M, Banka S, Moulton T, Curtin J, Rezaei N, Corns J, Innis JW, Avci Z, Chi Tran H, Pellier I, Pierani P, Fruge R, Parvaneh N, Mamishi S, Mody R, Darbyshire P, Motwani J, Murray J, Buchanan GR, Newman WG, Alter BP, Boxer LA, Donadieu J, Welte K, Klein C: **Extended Spectrum of Human Glucose-6-Phosphatase Catalytic Subunit 3 Deficiency: novel Genotypes and Phenotypic Variability in Severe Congenital Neutropenia.** *J Pediatr* 2011, **160**:679–683.
12. Lin JH, Walter P, Yen TS: **Endoplasmic reticulum stress in disease pathogenesis.** *Annu Rev Pathol* 2008, **3**:399–425.
13. Mc Dermott DH, De Ravin SS, Sik Jun H, Liu Q, Long Priel DA, Noel P, Takemoto CM, Ojode T, Paul SM, Dunsmore KP, Hilligoss D, Marquesen M, Ulrick J, Kuhns DB, Chou JY, Malech HL, Murphy PM: **Severe congenital**

- neutropenia resulting from G6PC3 deficiency with increased neutrophil CXCR4 expression and myelokathexis. *Blood* 2010, **116**:2793–2802.
14. Banka S, Wynn R, Newman WG: **Variability of bone marrow morphology in G6PC3 mutations: Is there a genotype-phenotype correlation or age dependent relationship?** *Am J Hematol* 2011, **86**(2):235–237.
  15. Banka S, Chervinsky E, Newman WG, Crow YJ, Yeganeh S, Yacobovich J, Donnai D, Shalev S: **Further delineation of the phenotype of severe congenital neutropenia type 4 due to mutations in G6PC3.** *Eur J Hum Genet* 2011, **19**:18–22.
  16. Germeshausen M, Zeidler C, Stuhmann M, Lanciotti M, Ballmaier M, Welte K: **Digenic mutations in severe congenital neutropenia.** *Haematologica* 2010, **95**:1207–1210.
  17. Yeshayahu Y, Asaf R, Dubnov-Raz G, Schiby G, Simon AJ, Lev A, Somech R: **Testicular failure in a patient with G6PC3 deficiency.** *Pediatr Res* 2014. doi:10.1038/pr.2014.64.
  18. Bonilla MA, Gillio AP, Ruggeiro M, Kernan NA, Brochstein JA, Abboud M, Fumagalli L, Vincent M, Gabrilove JL, Welte K, Souza LM, O'Reilly RJ: **Effects of recombinant human granulocyte colony-stimulating factor on neutropenia in patients with congenital agranulocytosis.** *N Engl J Med* 1989, **320**:1574–1580.
  19. Dale DC, Bonilla MA, Davis MW, Nakanishi AM, Hammond WP, Kurtzberg J, Wang W, Jakubowski A, Winton E, Lalezari P: **A randomized controlled phase III trial of recombinant human granulocyte colony-stimulating factor (filgrastim) for treatment of severe chronic neutropenia.** *Blood* 1993, **81**:2496–2502.
  20. Donadieu J, Leblanc T, Bade Meunier B, Barkaoui M, Fenneteau O, Bertrand Y, Maier-Redelsperger M, Micheau M, Stephan JL, Phillipe N, Bordigoni P, Babin-Boilletot A, Bensaid P, Manel AM, Vilmer E, Thuret I, Blanche S, Gluckman E, Fischer A, Mechinaud F, Joly B, Lamy T, Hermine O, Cassinat B, Bellanné-Chantelot C, Chomienne C on behalf the French Severe Chronic Neutropenia study group: **Analysis of risk factors for myelodysplasias, leukemias and death from infection among patients with congenital neutropenia. Experience of the French Severe Chronic Neutropenia Study Group.** *Haematologica* 2005, **90**:45–53.

doi:10.1186/s13052-014-0080-8

**Cite this article as:** Notarangelo et al.: Severe congenital neutropenia due to G6PC3 deficiency: early and delayed phenotype in two patients with two novel mutations. *Italian Journal of Pediatrics* 2014 **40**:80.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

